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Synthesis and Reactivity of Novel Spirocarbocycles as Scaffolds for New Nucleoside Analogues

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Abstract

A novel class of substituted spiro[3.4]octanes can be accessed via a [2+2]-cycloaddition of dichloroketene on a readily prepared *exo*-methylene cyclopentane building block. This reaction sequence was found to be robust on multigram scale and afforded a central spirocyclobutanone scaffold for carbocyclic nucleosides. The reactivity of this constrained building block was evaluated and compared to the corresponding 4'-spirocyclic furanose analogues. Density functional theory (DFT) calculations were performed to support the observed selectivity in the carbonyl reduction of spirocyclobutanone building blocks. Starting from novel spirocyclic intermediates, we exemplified the preparation of an undescribed class of carbocyclic nucleoside analogues and provided a proof of concept for the application as inhibitors for the protein methyltransferase target PRMT5.

Introduction

The term carbocyclic nucleoside is used to refer to structural analogues of naturally occurring and synthetic nucleosides in which the oxygen atom of the sugar ring is replaced by carbon. These nucleoside isosteres are characterized by an increased metabolic stability when compared to the riboside analogues as they are not recognized by e.a. nucleoside phosphorylases and hydrolases that otherwise can cleave the glycosidic bond.^[1] Consequently, the synthetic design^[2] and biological properties^[3] of various carbocyclic nucleosides have been well documented. with applications of this compound class in the therapeutic areas of antiviral and anticancer drug discovery research. Important examples of carbocyclic nucleosides that have been marketed include the antiviral drug abacavir (1),^[4] which is a highly potent nucleoside reverse transcriptase inhibitor used as anti-HIV medication, and entecavir (2),^[5] used in the treatment of patients suffering from hepatitis B infection (Figure 1). In a recent example, carbocyclic analogue 3b of the anticancer drug decitabine (3a) was found to be significantly more stable towards hydrolysis while

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maintaining activity towards DNA methyltransferases.^[6] Also in the field of anticancer agents, the carbocyclic analogue **4b** of the potent DOT1L histone methyltransferase inhibitor pinometostat (**4a**) is currently undergoing clinical trials for the treatment of acute leukemia and has demonstrated an improved metabolic stability compared to **4a** while desirable target activity and selectivity is conserved.^[7] Furthermore, compound **5** showed to be a highly potent SAM-competitive methyltransferase inhibitor for PRMT5:MEP50 multimer complex (IC₅₀ = 0.14 nM), displaying a high selectivity and favourable pharmacokinetic profile in different *in vitro* and cellular assays, and is efficacious *in vivo* in certain tumor xenograft models.^[8]

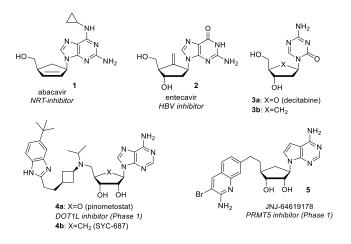
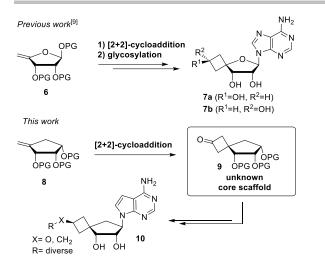


Figure 1. Examples of carbocyclic nucleoside analogues in medicinal chemistry.

As depicted in Scheme 1, we previously developed the synthesis of conformationally constrained 4'-spirocyclobutyl nucleoside analogues (type 7) using a [2+2]-cycloaddition of dichloroketene on 4'-exo-methylene furanose substrates (type 6), followed by Vorbrüggen nucleobase introduction.^[9] However, a synthetic method that includes the corresponding carbocyclic analogues of 7 has not been described to date. The development of a strategy that allows access to carbocyclic core scaffolds of type 9 would result in a valuable expansion with the potential to discover biologically active constrained nucleoside analogues of type 10. Indeed, building block 9 can serve as a modular scaffold in the preparation of carbocyclic nucleoside mimetics, as well as for the introduction of various pharmacophore substituents at the cyclobutanone moiety. Replacing the endocyclic furanose oxygen in 6 by a carbon atom, however, is expected to alter the reactivity of the double bond, so that the question whether in this case a similar cycloaddition strategy would be possible and hence, whether spirocarbocyclic nucleoside 10 can be accessed, remained to be investigated.



Scheme 1. Synthesis of 4'-spirocyclic nucleoside analogues of type **7** (previous work^[9]) and envisioned preparation of carbocyclic nucleoside analogues of type **10** (this work)

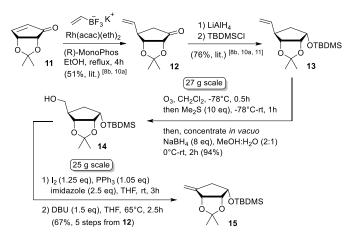
Results and Discussion

Commercially available cyclopentenone **11**, a frequently used synthon in the synthesis of carbocyclic nucleoside analogues,^[10] was used as a chiral building block in the preparation of *exo*-methylene substrate **15**. The conversion of **11** towards alcohol **14** has been reported in the literature, starting with a Michael addition on cyclopentenone **11** via rhodium-catalysed addition of potassium vinyltrifluoroborate^[8b] or by using vinylmagnesium bromide as nucleophile,^[10a] affording substrate **12**. Sequentially, stereoselective reduction of ketone **12** and silylation of the resulting alcohol was performed to give cyclopentane **13** (Scheme 2) in 71% overall yield from **12**.^[8b, 10a, 11]

The oxidation of 13 to the intermediate aldehyde has been described using catalytic amounts of OsO4 in combination with cooxidant NaIO₄, followed by reduction to give the corresponding alcohol (14).^[11] However, when we used these oxidation conditions, inconsistent conversions from the starting material (13) were observed on small scale (0.5-1.0 mmol), and the formation of side products was noticed. Aiming to improve the reproducibility of the reaction on multigram scale, we evaluated the oxidation of 13 using ozone as a milder alternative that does not require the use of transition metals and simplifies reaction work-up. To facilitate this, ozone was electrochemically generated from oxygen and bubbled through a solution of 13 in dichloromethane at -78°C. Applying these reaction conditions, we observed complete conversion from the starting material (13) and upon treatment of the intermediate aldehyde with an excess of NaBH₄ after a solvent switch to aqueous methanol, the desired alcohol (14) was obtained in high yield (Scheme 2).

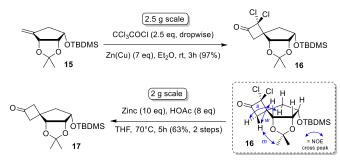
To obtain key exo-methylene intermediates of type 8 (Scheme 1), an Appel reaction was performed on alcohol 14 and the iodomethyl intermediate was treated with DBU in THF to afford 15 (Scheme 2). The synthesis of building block 15 was successfully repeated on a 25 gram scale in an excellent overall yield of 67% starting from ketone 12, requiring only a single purification via

silica gel chromatography after the final elimination step towards alkene **15**.



Scheme 2. Synthesis of exo-methylene 15 from cyclopentenone 11

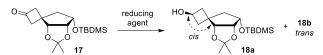
Having access to multigram quantities of building block 15, the [2+2]-cycloaddition reaction of dichloroketene was attempted using standard reaction conditions as performed previously on 4'exo-methylene substrates of type 6, i.e. the dropwise addition of trichloroacetyl chloride in anhydrous diethyl ether to a mixture of substrate 15 with activated zinc powder.^[9, 12] We were pleased to observe that by using this protocol, cycloadduct 16 was obtained as a single diastereoisomer in an almost quantitative amount (Scheme 3). The structure of 16 was elucidated via NOE NMR analysis, which confirmed that dichloroketene reacts with exomethylene substrate 15 from the least sterically hindered β-face (Scheme 3). It was noticed that the purification of 16 via normal phase silica gel chromatography was not efficient and resulted in a low recovery, presumably due to degradation via ring opening of the dichlorocyclobutanone. Given the high purity in which 16 was obtained after the [2+2]-cycloaddition reaction, no further attempts were made to purify the cycloadduct (16) at this stage. Reductive dechlorination was performed via treatment of 16 with zinc and acetic acid, which readily afforded the stable cyclobutanone product 17 in good overall yield after purification via silica gel chromatography (Scheme 3).



Scheme 3. [2+2]-Cycloaddition of dichloroketene on *exo*-methylene 15 and reductive dechlorination towards cyclobutanone 17.

Next, the selectivity of the carbonyl reduction was investigated for spirocyclobutanone **17**. An initial screening with commonly used reducing agents, including NaBH₄, LiBH₄ or LiAlH₄, all resulted in the formation of 1:1 mixtures of alcohols **18a** (*cis*) and **18b** (*trans*). As depicted in Table 1, only a slight preference for *cis* isomer **18a** (56%) was observed when L-selectride (entry 4) was

used as reducing agent. A modest selectivity was noticed when borane reductions were used in combination with chiral Corey-Itsuno (Corey-Bakshi-Shibata, CBS) oxazaborolidine catalysts.^[13] Here, a preference of 67% for *cis* alcohol **18a** (entry 5, Table 1) and 58% for *trans* alcohol **18b** (entry 6) could be obtained at best, depending on the chirality of the CBS catalyst used. Apart from the rather moderate selectivity, longer reaction times were required and generally incomplete conversion from the starting material (**17**) was obtained. These results, together with the observation that diastereomers **18a** and **18b** could not easily be separated via normal phase silica gel chromatography or preparative HPLC separation but required the use of supercriticalfluid chromatography (SFC) instead, showed that this method of reduction is of limited synthetic value.



Entry	Reducing agent	Solvent	Time /h	Conv. /%	Ratio 18a : 18b
1	NaBH ₄	MeOH	1	100	50:50
2	LiBH ₄	THF	1.5	100	50:50
3	LiAIH ₄	THF	1.5	100	50:50
4	L-selectride	THF	1.5	100	56:44
5	BH ₃ /(R)-MeCBS	THF	20	64	67:33
6	BH ₃ /(S)-MeCBS	THF	20	100	42:58
7	BH ₃ /(<i>R</i>)-BuCBS	THF	21	52	67:33
8	BH ₃ /(S)-BuCBS	THF	21	51	42:58
9	BH₃/(<i>R</i>)- <i>o</i> -tolyl CBS	THF	21	31	60:40
10	BH₃/(<i>S</i>)- <i>o</i> -tolyl CBS	THF	21	34	50:50

Table 1. Screening of reducing agents on cyclobutanone 17 (0.15 mmol scale)

The results listed in Table 1 are in sharp contrast when compared to previous carbonyl reductions on ribose derived cyclobutanones 19 and 20 (Scheme 4), where a high selectivity for both alcohol 21a (94%, cis) and 22a (88%, cis) was observed.^[9] We hypothesized that this selectivity could be attributed to a repulsive electrostatic interaction of the incoming hydride with the endocyclic furanose oxygen, which directs the approach to the opposite (trans) side, leading to a preferred formation of cis isomers 21a and 22a. These results are indeed in agreement with a literature precedent on the reduction of 3alkoxycyclobutanones, affording cis 3-alkoxycyclobutanols as major reaction products (9:1 ratio).^[14] Since this electrostatic repulsive effect does not exist for cyclopentane analogue 17, and because no additional steric differences are present at both faces of the cyclobutanone, no significant selectivity is expected in the carbonyl reduction of 17. To validate our hypothesis for the significant selectivity differences in the carbonyl reductions of substrates 17 and 19, density functional theory (DFT) calculations were performed.

Based on our previous computational study, [15] the reduction of 4'-spirocyclobutanones with LiAlH4 was described using four

different transition states in which the lithium counterion coordinates either to one (bidentate) or two hydrogen atoms (tridentate). The cis alcohol products of both reductions (18a and 21a) always arise from a pathway involving the bidentate TS_{cis} transition state or its tridentate analogue TS'cis, while the remaining transition states TS_{trans} and TS'_{trans} yield the trans isomers 18b and 21b. Consequently, the cis isomer 18a is obtained from a syn-facial approach of the reducing agent with respect to the cyclopentane ring of compound 17, while the trans cyclobutanol 18b results from an anti-face attack via TStrans and TS' trans. In the case of the ribose derived spirocyclobutanone 19, the opposite face approach of LiAIH₄ relative to the endocyclic furanose oxygen atom yields cis cyclobutanol 21a, whereas alcohol 21b arises from the syn-facial hydride attack. The calculated Gibbs free reaction profiles for these competing transition states (Figure 1) indicate that the pathway involving the bidentate transition state $\mathbf{TS}_{\mathit{cis}}$ exhibits the lowest energy barrier for both 4'-spirocyclobutanones and hence, the cis cyclobutanols 18a and 21a should be obtained as major isomer. The syn-facial approach of the hydride towards cyclopentane analogue 17 involves an energy barrier of 11.0 kcal mol⁻¹, while the computed activation barrier for the reduction of 19 through TS_{cis} is 10.0 kcal mol⁻¹ at 195.15 K. However, the bidentate TS_{trans} pathways resulting in isomers 18b and 21b have activation barriers that are only 0.2 and 0.8 kcal mol⁻¹, respectively, higher than the most favorable pathways. On the other hand, the activation barriers involving tridentate transition states turn out to be systematically 1 to 2 kcal mol⁻¹ higher in energy than the bidentate pathways, though the preference for the *cis* isomer is still retained. Despite the preference for **TS**_{cis} pathways, a clear difference in selectivity between both reductions is noted from the estimated Boltzmann populations. While the reduction of spirocyclobutanone 19 yields 89% of cis product 21a, which is in close agreement with the experimental value (94%), the reduction of compound 17 results in 63% of cis alcohol. These findings fully support the previously mentioned experimental observations.

To assess the driving forces behind the distinct stereoselectivity of these reductions, we have analyzed the structural changes in the cyclobutanone ring arising from the hydride approach. The puckering angle (ϕ), defined as the angle formed by the intersection between the C2-C1-C4 and C2-C3-C4 planes,^[16] and the dihedral angle $D(O-C_1-C_2-H'_2)$ are two geometrical parameters that allow to quantify, respectively, the torsional strain and eclipsed interactions relative to the initial reagents (Figure S1). On one side, an increase in ϕ with respect to the reference values indicates a relief in torsional strain, while the cyclobutanone ring becomes more puckered. On the other side, a decrease in D denotes less eclipsed interactions between the carbonyl bond and the neighboring C-H bond. Both parameters have been previously used to establish the role of torsional effects on the stereoselectivity of cyclobutanone reductions.^[15,16] The relative Gibbs free energies and structural parameters for the different transition states together with the reference values for the reactants 17 and 19 are summarized in Table 2.

From Table 2, it is clear that both *anti*- and *syn*-facial attacks of the hydride involve a reduction in torsional strain, since ϕ increases with respect to the reactants, except for **TS**'_{*trans*} of **19**. In the case of spirocyclobutanone **19**, the preference for **TS**_{*cis*} can be attributed to the largest increase in ring puckering ($\Delta \phi = +3.6^{\circ}$) as well as the relief of eclipsed interactions ($\Delta D = -9.2^{\circ}$). By contrast, the preference for the reduction pathway involving **TS**_{*cis*} for **17** cannot be ascribed to torsional effects, since the *anti*-facial attack via **TS**'*trans* and **TS**'*trans* entangles a larger increase in puckering angle than the *syn*-facial hydride approach. Consequently, we further investigated the role of the noncovalent interactions using the noncovalent interaction (NCI) index.^[17]

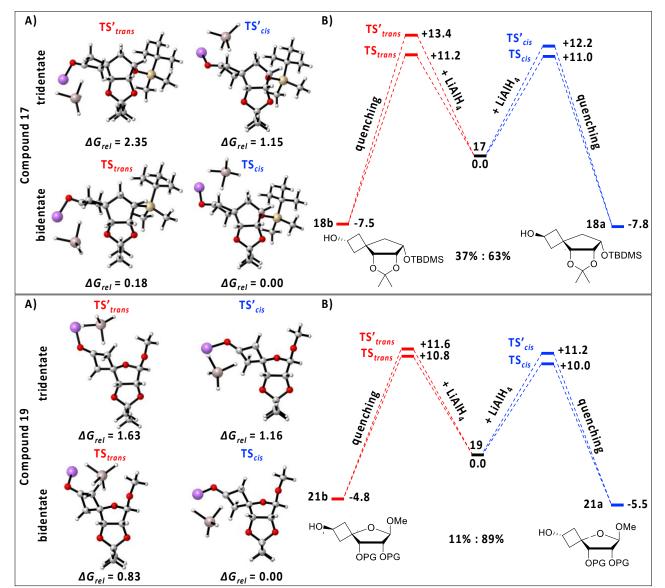


Figure 1. A) Optimized geometries for the various transition states involved in the reduction of the 4'-spirocyclobutanones 17 and 19 with LiAIH₄. B) Gibbs free reaction profiles (in kcal mol⁻¹) for the different pathways considered in the hydride reduction of 17 and 19 in THF at 195.15 K. The quenching process refers to the addition of water and subsequent removal of the reducing agent.

	Compound 17					Compound 19				
	ref ^[a]	TS _{cis}	TS _{trans}	TS'cis	TS'trans	ref ^[a]	TS cis	TS _{trans}	TS'cis	TS'trans
ΔG_{rel}	-	0.00	0.18	1.15	2.35	-	0.00	0.83	1.16	1.63
φ	14.2	15.2	17.9	16.6	19.7	15.8	19.4	17.0	19.0	15.5
Ď	79.3	80.9	72.1	82.2	73.8	82.8	73.6	77.4	77.2	83.1

Table 2. Relative Gibbs free energies (in kcal mol⁻¹) calculated in THF at 195.15 K together with selected torsional parameters (ϕ and *D* in °) for the different transition states involved in the reduction of the 4'-spirocyclobutanones **17** and **19** with LiAlH₄. ^[a] The reference values correspond to the cyclobutanone ring in the reactant structures of **17** and **19**.

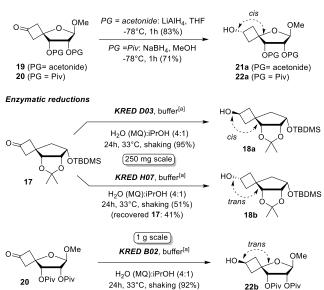
The two-dimensional plots of the reduced density gradient (s) with respect to the electron density (ρ) multiplied by the sign of the second eigenvalue of the electron density Hessian matrix (λ_2) allow to discern attractive interactions ($\lambda_2 < 0$) from repulsive ($\lambda_2 > 0$) ones. Furthermore, the visualization of the NCIs in real space is possible by representing the reduced gradient isosurfaces in three dimensions using a RGB color scale, with red isosurfaces corresponding to repulsive interactions, green ones to weak van der Waals interactions and attractive interactions appearing as blue isosurfaces. The isosurfaces of the four different transition states involved in the reduction of compound **17** and **19** are respectively shown in Figures **S2** and **S3**. The NCI plots (Figure **S3**) show a repulsive electrostatic interaction (marked with a circle) involving the endocyclic furanose oxygen atom that

hampers the *syn*-facial approach of LiAlH₄ towards the ribose derived spirocyclobutanone **19** via **TS**_{trans} and **TS**'_{trans}. It can therefore be concluded that the pronounced stereoselectivity for the *cis* cyclobutanol **21a** is driven by both the relief of repulsive interactions of the incoming hydride with the furanose oxygen atom and the largest decrease in torsional strain. On the other hand, repulsive interactions are present for both the *syn*- and *anti*facial approach of the hydride towards spirocyclobutanone **17**, though these interactions are more pronounced between the acetate protective group and the *anti*-facially attacking hydride in **TS**_{trans}(red circle in Figure **S2**). Consequently, we hypothesize that the poor selectivity of the reduction of **17** is due to a competition between a hydride approach delivering maximal torsional strain relief or minimal repulsive interactions.

To improve the poor stereoselectivity in the reduction of cyclobutanone 17, we investigated biocatalytic ketone reductions, as various ketoreductases (KREDs) are commercially available and these enzymes are typically characterized by a broad substrate scope.^[18] To this end, a screening kit ^[19] containing 19 engineered KRED enzymes derived from Lactobacillus kefir and Lactobacillus brevis wild-type enzymes was evaluated. A smallscale screening using the different enzymes on spirocyclobutanone substrate 17 revealed five ketoreductases which showed an excellent selectivity exceeding 95% for both diastereoisomers 18a and 18b, combined with high conversions from ketone substrate 17, as depicted in Table 3 (entries 1-5). Consequently, the enzymes that displayed the most promising outcome based on small-scale screening were selected to evaluate the stereoselective synthesis of alcohols 18a (cis) and 18b (trans). Cyclobutanone 17 dissolved in isopropyl alcohol was mixed with enzyme D03 (entry 3. Table 3) and the aqueous buffer solution containing the cofactor was moderately shaken for 24 hours at 33°C. After a short elution over silica gel column, we observed that alcohol 18a was obtained as a single isomer in excellent isolated yield (95%) (Scheme 4). Likewise, alcohol 18b (trans) could be obtained in a diastereoselective way using a different ketoreductase (entry 5, Table 3). Due to the partial conversion in the latter reduction, 18b was obtained in a lower yield, however, the unreacted ketone (17) could be efficiently recovered via silica gel chromatography.

The successful stereoselective transformation of ketone 17 to alcohols 18a (cis) and 18b (trans) prompted us to repeat the screening with the different KREDs on ribose derived cyclobutanone 20, where previously a maximum chemoselectivity of 88% for isomer 22a (cis) was obtained using hydride reducing agents (Scheme 4). Also in this case, several enzymes were identified for the diastereoselective reduction of spirocyclobutanone 20 to alcohol 22b (trans), of which a selection is listed in Table 3 (entries 6-9). Encouraged by these results, the enzyme that resulted in a diastereoselective reduction combined with the highest conversion from ketone 20 (entry 8, Table 3) was used for scale-up, which resulted in the development of a reproducible, high yielding protocol to transform ketone 20 to alcohol 22b (trans) on one-gram scale. Consequently, this biocatalytic reduction using a green solvent mixture became the preferred method to obtain alcohol 22b (trans), thereby avoiding the cumbersome isolation of this isomer from a cis-trans mixture of diastereoisomers 22a and 22b. Noteworthy, only one enzyme (entry 9, Table 3) was found to be partly (90%) selective for alcohol 22a (cis), previously identified as the major reaction product in the screening of chemical carbonyl reductions (Scheme 4). Overall, this enzymatic reduction protocol affords a highly efficient approach for the stereoselective synthesis of alcohols 18a, 18b and 22b, making use of commercially available ketoreductases.

Previous results

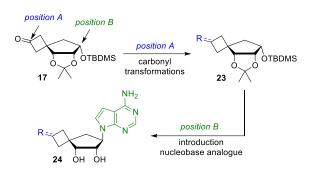


Scheme 4. Carbonyl reductions of cyclobutanones 17 and 19-20, [a] buffer: Na₃PO₄ (128 mM), MgSO₄ (1.7 mM), NADP⁺ (1.1 mM) at pH 7.0 dissolved in H₂O (MQ)

Entry	KRED	ketone	Conv. /%	Ratio a : b ^[a]
1	B05	17	100	95:5
2	B02	17	80	≥ 99% 18a
3	D03	17	100	≥ 99% 18a
4	A12	17	95	3:97
5	H07	17	77	≥ 99% 18b
6	C01	20	65	2:98
7	C02	20	73	≥ 99% 22b
8	B02	20	93	≥ 99% 22b
9	A12	20	12	90:10

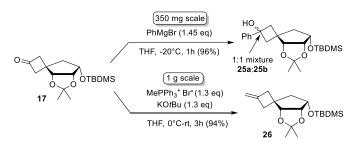
Table 3. Screening enzymatic carbonyl reductions of cyclobutanones 17 and 20, [a] 18a:18b (substrate 17) or 22a:22b (substrate 20)

Having established a robust protocol towards the central carba-nucleoside core scaffold **17**, we embarked on the introduction of functional group handles and key pharmacophores at two relevant positions in **17** (position A and B, Scheme 5). Specifically, we attempted to 1) introduce a tertiary phenyl carbinol (towards a constrained mimetic of the methyltransferase inhibitor LLY-283^[20]); 2) introduce an exocyclic alkene synthesis of spirocyclic analogues of PRMT5 inhibitor **5**^[8a-c]); and 3) to install a nucleobase analogue at the relevant position towards compounds of type **24**, as depicted in Scheme **5**. Moreover, we investigated the differences in chemical reactivity between the novel spirocarbocyclic building block **17** and the corresponding furanose analogues **19** and **20**.



Scheme 5. Functionalization of core scaffold 17

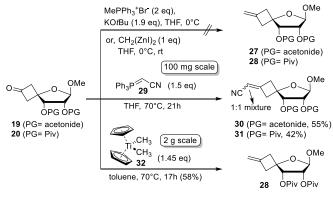
The reactivity evaluation of ketone 17 was initiated by treatment with phenylmagnesium bromide at low temperatures, which readily afforded alcohols 25a and 25b in a 1:1 ratio (Scheme 6). This Grignard addition displayed no stereoselectivity, similarly to the carbonyl reduction of 17 as listed in Table 1. Nonetheless, both tertiary alcohols 25a and 25b could be separated in an efficient manner via SFC purification and were obtained in good isolated yields. As stated previously, these intermediates could be used as building blocks for the preparation of constrained, carbocyclic analogues of the PRMT5 inhibitor LLY-283.^[20] To further explore the synthetic potential of ketone 17, a Wittig reaction was attempted in order to prepare exo-methylene 26 (Scheme 6). Therefore, methyltriphenyl phosphonium bromide was treated with KOtBu to generate the corresponding Wittig ylide in situ, which was then added in portions to a solution of ketone 17 in THF. This reaction efficiently afforded alkene 26 in excellent isolated yield after a short elution over silica gel column. The terminal alkene moiety in 26 can serve as a useful functional handle for further derivatization via e.g. hydroboration reactions or transition metal catalysed cross couplings.



Scheme 6. Grignard addition and Wittig reaction on cyclobutanone 17

Surprisingly, when the same Wittig reaction was attempted on ribose analogues **19** and **20** to prepare alkenes **27** and **28**, respectively, an instantaneous degradation of the ketones was observed during the addition of the Wittig ylide, resulting in the formation of a dark-yellow coloured mixture (Scheme 7). The instability of the latter ketone substrates under basic reaction conditions was confirmed by a series of ¹H NMR monitorred experiments in which different bases (KO*t*Bu, DBU and NaH) were added to **19** and **20** in THF, showing a fast decomposition. It is very likely that ketones **19** and **20** are readily enolized at the cyclobutanone when subjected to a strong base and undergo fragmentation through ring opening of the furanose ring. This hypothesis was further confirmed when the deactivated and less basic phosphonium ylide **29** was found to successfully convert ketones **19** and **20** to alkenes **30** and **31**, resulting in isomeric mixtures on both substrates (Scheme 7). The instability of **19** and **20** towards basic reaction conditions clearly does not apply to cyclopentane analogue **17**, as both Grignard addition and Wittig reaction are compatible and afford the desired products in high yields (Scheme 6). The absence of the endocyclic oxygen atom in ketone **17** thus prevents the decomposition of the spirocyclic scaffold after α -deprotonation of the cyclobutanone.

In order to synthesize alkene 28, an alternatively approach was followed that uses bis(iodozincio)methane reagent, which is reported to have a reduced basicity as compared to the previously employed Wittig ylide (Scheme 7).[21] Unfortunately, no conversion was observed from ketones 19 or 20 towards the desired products, and decomposition of the starting material was observed instead. In a final attempt, the Petasis reagent (32) was tested to facilitate the olefination of ketone 20 (Scheme 7).[22] To this end, commercially available dimethyltitanocene 32 was mixed with ketone 20 in toluene and heated to 70°C to generate the active methylene species in-situ. Gratifyingly, this reaction successfullv resulted in а clean conversion from spirocyclobutanone 20 towards the desired alkene 28, which was isolated in 58% yield on two-gram scale after silica gel chromatography.

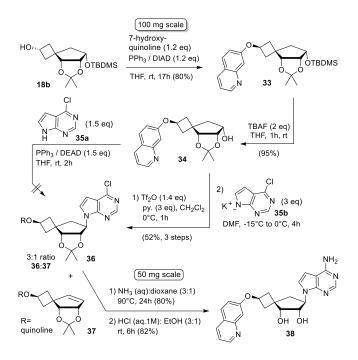


Scheme 7. Evaluation of olefination reactions on cyclobutanones 19 and 20

Next, we evaluated if the obtained novel spirocarbocyclic scaffold structures can be used as building blocks for the preparation of the corresponding nucleoside analogues. To deliver a proof of concept for this transformation, alcohol 18b was selected as a substrate in the synthesis of pyrrolopyrimidine nucleoside analogue 38 (Scheme 8). In analogy to the potent methyltransferase inhibitor 5 (Figure 1), we planned the introduction of a quinoline pharmacophore at the cyclobutyl alcohol, as this can potentially result in biologically active nucleoside analogues as mimetics of S-adenosylmethionine (SAM). Therefore, prior to the introduction of the pyrrolopyrimidine moiety, the cyclobutyl alcohol 18b was transformed to 33 via a Mitsunobu reaction with inversion of stereochemistry. Subsequently, the treatment of 33 with tetrabutylammonium fluoride (TBAF) efficiently removed the silvl protecting group to afford alcohol 34 as a suitable substrate for the introduction of a heterocyclic pyrrolopyrimidine base. Using 6-chloro-7deazapurine (35a) as reagent, initial attempts were made to introduce this heterocyclic moiety via a Mitsunobu reaction on alcohol 34 (Scheme 8), however, this approach turned out to be

not synthetically useful and resulted in the formation of numerous side products. Therefore, this strategy was not investigated further, and focus was directed to the nucleophilic substitution with deazapurine **35b** on O-triflated substrate **34**, as described for related cyclopentanols.^[8c,23] First, **34** was treated with triflic anhydride to prepare the intermediate triflate, followed by a nucleophilic substitution using an excess of preformed potassium salt **35b**. A complete conversion towards desired product **36** was observed after 4 hours, alongside a fraction of elimination side product **37** in a 3:1 ratio, respectively. After an efficient removal of **37** via silica gel chromatography, the desired intermediate (**36**) was isolated in a 52% overall yield from **33**.

Finally, substitution of the chlorine atom in **36** was carried out via treatment with aqueous ammonia in dioxane at elevated temperature, followed by acidic hydrolysis using a mixture of aqueous HCI in ethanol to deprotect the acetal group. This reaction sequence successfully afforded carbocyclic nucleoside analogue **38** in good overall yield (Scheme 8). This example validates the compatibility of the synthetic methodology to access a novel class of pharmacologically relevant carbospirocyclic nucleoside analogues.

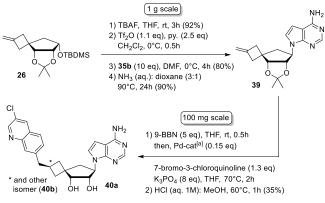


Scheme 8. Synthesis of carbocyclic nucleoside analogue 38 from spirocyclic building block 18b

To demonstrate that *exo*-methylene **26** offers a useful handle for the late stage functionalization of the cyclobutyl ring, we attempted the preparation of the derived nucleoside analogue **39** (Scheme 9). To facilitate this, the silyl protecting group in **26** was removed, followed by treatment with triflic anhydride and substitution of the triflate with pyrrolopyrimidine **35b**, which showed an improved selectivity for the desired substitution product over elimination side product (82:18 ratio, respectively). In analogy to substrate **36** (Scheme 8), treatment of the corresponding intermediate with ammonia successfully afforded building block **39** in an excellent overall yield. We were pleased to find that the treatment of **39** with 9-BBN readily afforded the

intermediate borane and could successfully be converted towards the corresponding cross-coupled products as exemplified by the introduction of a 3-chloroquinoline moiety, which is a highly potent PRMT5 pharmacophore, at the cyclobutyl ring. ^[24] As depicted in Scheme 9, final deprotection of the acetonide allowed the isolation of spirocarbocyclic nucleoside analogues **40a** and **40b** in a 1:1 ratio. The above synthetic sequence thus demonstrates to be compatible with the novel spirocarbocyclic nucleoside building blocks of type **39**, enabling access to a variety of constrained mimetics of the potent PRMT5 inhibitor **5**.

The examination of the in vitro activity of novel 4'spirocarbocyclic nucleoside analogues 40a and 40b in the PRMT5/MEP50 multimer complex indicates a promising micromolar inhibition of 2.00 and 1.09 µM (IC₅₀ values), respectively. Furthermore, we observed that compound 38 (Scheme 8), bearing a non-functionalized auinoline pharmacophore attached to the cyclobutyl moiety via an oxygen linker, resulted in a 10-fold increase in potency compared to 40b, showing a sub-micromolar inhibition value of 0.09 µM in the PRMT5/MEP50 assay. These biological results clearly indicate that novel spirocarbocycles of types 18 and 26 are valuable building blocks for the synthesis of a variety of potential competitive PRMT5 inhibitors.



Scheme 9. Synthesis of carbocyclic nucleoside analogues 40a and 40b from spirocyclic building block 26, [a] Pd-cat: 1,1'-bis(di-*tert*-butylphosphino)-ferrocene palladium dichloride

Conclusions

We have achieved the synthesis of a novel carbospirocyclic core scaffold (17) on multigram scale, starting from commercially available cyclopentenone 11, and demonstrated that derived nucleosides analogues could be accessed from this key building block. *Exo*-methylene 15 was found to be an excellent substrate for the [2+2]-cycloaddition of dichloroketene, affording cycloadduct 16 in a stereoselective way. An evaluation of the reactivity of the derived cyclobutanone (17) revealed significant differences when compared to previously prepared spirocyclic ribose analogues 19 and 20. We supported the experimentally observed differences in carbonyl reduction selectivity between substrates 17 and 19 by means of DFT calculations and found that chemoenzymatic reductions of ketone 17 were successful to selectively prepare a single stereoisomer (18a or 18b), whereas

conventional carbonyl reductions proved to be significantly inferior. Also, cyclobutanone **17** was found to be compatible with conditions involving basic reagents, such as Grignard and Wittig reactions. In contrast, ketones **19** and **20** derived from ribose were unstable under such conditions, requiring a different synthetic approach. We showed that core scaffold **17** can be efficiently transformed into carbospirocyclic nucleoside analogues, either after derivatizations of the cyclobutanone functional group handle, or prior to the introduction of desired pharmacophore groups, allowing late stage functionalization of intermediate **39**. Finally, we delivered a proof of concept that spirocarbocycles of type **18** and **26** are modular building blocks for the preparation of potent inhibitors for methyltransferase PRMT5.

Experimental section

Materials and instrumentation. Nuclear magnetic resonance (NMR). For synthesized compounds, ¹H NMR and ¹³C NMR spectra were recorded on a Bruker DPX-360 operating at 360 MHz for ¹H NMR and 91 MHz for ¹³C NMR, on a Bruker Avance 400 operating at 400 MHz for ¹H NMR and 101 MHz for ¹³C NMR, or on a Bruker Avance III 400 operating at 400 MHz for ¹H NMR and 101 MHz for ¹³C NMR. Alternatively, ¹H and ¹³C NMR spectra were recorded at 500 MHz for ¹H NMR and 125 MHz for ¹³C NMR on a Bruker Avance II 500 console, or at 250 MHz for ¹H NMR and 63 MHz for ¹³C NMR on a Bruker Avance DRX 250 console. Tetramethylsilane (TMS) was used as an internal standard. The used solvent and frequency are mentioned before every analysis. Chemical shifts (δ) are reported in parts per million (ppm) relative to tetramethylsilane (TMS). Coupling constants (J) are given in Hertz (Hz). The following abbreviations are used in the description of spectra: singlet (s), doublet (d), triplet (t), quadruplet (q), quintet (qn), multiplet (m), doublet of doublets (dd), triplet of doublets (td), doublet of triplets (dt), doublet of doublet of doublets (ddd), pseudo (ps). Liquid Chromatography - mass spectroscopy (LC-MS). High Performance Liquid Chromatography (HPLC) measurement was performed using a LC pump, a diode-array (DAD) UV detector and a column as specified in the respective methods (Entries 1-4, Table S1, Supporting Information). Flow from the column was brought to the Mass Spectrometer (MS) which was configured with an atmospheric pressure ion source. Data acquisition was performed with MassLynx software from Waters. Intermediates and final compounds are described by their experimental retention times (Rt) and ions. Electrospray ionization (ES+ and/or ES-) was used as ionization method. If not specified differently in the table of data, the reported molecular ion corresponds to the [M+H]⁺ (protonated molecule) and/or [M-H]-(deprotonated molecule). In case the compound was not directly ionizable the type of adduct is specified (i.e. [M+NH4]+, [M+HCOO]⁻, etc...). For molecules with multiple isotopic patterns (Br, Cl), the values are reported for the different isotopes with corresponding intensities, or the reported value is the one obtained for the lowest monoisotopic mass. Gas Chromatography - Mass Spectroscopy (GC-MS). GC measurement was carried out on a J&W HP-5MS column (20 m x 0.250 mm, 0.25 mm) from Agilent Technologies (instrument: GC6890- MSD5973N), with a constant pressure of 12.97 psi. The temperature gradient applied was as follows: initial temperature 50°C, hold for 0.1 min, then a 24°C/min ramp applied for 9.58 min until 280°C and hold for 5.0 min in a 14.68 min run. Front inlet temperature was 250°C. Split injection mode was used, 1 ml injection volume, with a 10/1 ratio into the GC-MS system and a flow of 2.3 ml/min. Electron ionisation was used as ionisation mode. Supercritical Fluid Chromatography -Mass Spectrometry (SFC-MS). SFC purifications were performed on an Acquity UPC2 system from Waters equipped with a UV, QDA and ELSD detector. The SFC measurement was performed using an Analytical Supercritical fluid chromatography (SFC) system composed by a binary pump for delivering carbon dioxide (CO₂) and modifier, an autosampler, a column oven, a diode array detector equipped with a high-pressure flow cell standing up to 400 bars. If configured with a Mass Spectrometer (MS) the flow from the column was brought to the (MS). Data acquisition was performed with MassLynx software from Waters. The analytical method used for SFC-MS is detailed in Table S2 (Supporting Information). High-Resolution Mass Spectrometry (HR-MS). UHPLC-HRMS analyses were performed using a Dionex Ultimate 3000 UHPLC system coupled to a Thermo Orbitrap Fusion Lumos Tribrid mass spectrometer. High resolution accurate mass data were acquired in positive ion mode using an electrospray ionization source. Note: For the following new molecules listed, HRMS data could not be provided as a result of poor ionization: 15-17, 18a, 18b, 25a, 25b, 26, 28, 30, 31 & 42. The identity of aformentioned compounds has been unambigiously confirmed by NMR analysis (see below). For all other intermediates prepared via linear synthesis strategy, HRMS data is included for at least every third compound in a row. This applies to intermediates 33, 37, 39, 41, & 43 (see experimental procedures and Silica characterization below). gel chromatography. Purifications were conducted manually using synthetic amorphous silica gel Silica gel 60 (Merck Millipore, 0.040 -0.063mm) or automatically using a Reveleris™ X2 Flash Chromatography System with integrated ELS/UV/UV-Vis detection and pre-packed silica gel cartridges (Reveleris™ SRC Silica Flash Cartridges, 40 µm) from Grace™ Reveleris™ or silica gel cartridges (puriFlash, 25 µm) from Interchim. Alternatively, a Biotage® SP4 Flash Chromatography system was used with integrated UV detection and pre-packed silica cartridges (SNAP ultra, 25 µm) from Biotage®.

Experimental procedures and characterization.

Synthesis (3aR,6R,6aR)-2,2-dimethyl-6of vinyltetrahydro-4H-cyclopenta[d][1,3]-dioxol-4-one 12.[8b] Acetylacetonatobis(ethylene)rhodium(I) (837 mg, 3.24 mmol, (R)-N,N-dimethyldinaphtho[2,1-d:1',2'-0.02 eq) and F][1,3,2]dioxaphosphepin-4-amine (2.91 g, 8.11 mmol, 0.05 eq) were dissolved in EtOH (625 ml) under nitrogen atmosphere. The mixture was stirred at room temperature and flushed with nitrogen for 15 minutes. Then, (-)-(3aR,6aR)-3a,6a-dihydro-2,2-dimethyl-4H-cyclopenta-1,3-dioxol-4-one (11, 25.0 g, 162.16 mmol, 1.00 eq) and potassium vinyltrifluoroborate (45.7 g, 324 mmol, 2.00 eq) were added and the reaction mixture was stirred at reflux for 4 hours. The mixture was cooled to room temperature, filtered over a path of celite and rinsed with *n*-heptane. A total volume of 500 ml of *n*-heptane was added, followed by water (300 ml) and the product was extracted in n-heptane (3x 500 ml). The combined organic layers were washed with NH₄OH (aq. 12%, 3x 250 ml) and brine (2x 250 ml). The organic layer was dried (MgSO₄), filtered and the solvent was removed by distillation at atmospheric pressure. n-Heptane (200 ml) was added to the residue and the solids were removed via decantation of the liquids. Further removal of residual solvent was performed via distillation at

atmospheric pressure to afford the title compound **12**. Dark red oil. Yield: 51% (16.2 g, crude). ¹H NMR (400 MHz, CDCl₃): δ 5.83 (ddd, J= 17.2, 10.7, 6.3 Hz, 1H), 5.05-5.20 (m, 2H), 4.64 (d, J= 4.9 Hz, 1H), 4.20 (d, J= 4.9 Hz, 1H), 3.07-3.15 (m, 1H), 2.84 (dd, J= 18.3, 8.5 Hz, 1H), 2.29 (br d, J= 18.3 Hz, 1H), 1.45 (s, 3H), 1.36 ppm (s, 3H). The spectroscopic results are consistent with reported literature data.^[8b, 10a, 11]

Synthesis of tert-butyl(((3aR,4S,6R,6aR)-2,2-dimethyl-6-vinyltetrahydro-4H-cyclopenta[d][1,3]dioxol-4-

yl)oxy)dimethylsilane 13.^[8b, 10a, 11, 24] A nitrogen flushed 4necked 1L flask was equipped with a pressure equalized dropping funnel, a thermometer, 2 septa and subsequently charged with THF (174 ml) and lithium aluminum hydride (5.71 g, 143 mmol, 1.50 eq), respectively. The mixture was cooled to 0°C and (3aR,6R,6aR)-2,2-dimethyl-6-vinyltetrahydro-4H-

cyclopenta[d][1,3]-dioxol-4-one 12 (17.4 g, 95.5 mmol, 1.00 eq) dissolved in THF (87 ml) was added dropwise to the mixture over a period of 1 hour during which a temperature between 0-5°C was maintained. After addition, the mixture was stirred for an additional 45 minutes at 0°C. Subsequently, the mixture was carefully quenched at 0°C with water (5.5 ml, dropwise over 15 min.) and NaOH (15% in H₂O, 5.5 ml) was added. The mixture was filtered over celite and rinsed with THF (300 ml). The mixture was concentrated in vacuo and the residue (16.6 g) was dissolved in pyridine (55 ml), cooled to 0°C and tert-butyldimethylsilyl chloride (14.9 g, 98.8 mmol, 1.00 eq) was added portion wise over 15 minutes. After addition, the mixture was warmed to room temperature and stirred for 22 hours. Subsequently, n-heptane (300 ml) was added to the solution followed by NaHCO₃ (ag. sat. 100 ml). The product was extracted in *n*-heptane (3x 300ml) and the combined organic layers were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: nheptane/EtOAc from 99:1 to 9:1). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 13. Slight yellow oil. Yield: 76% (19.8 g, 66.4 mmol). ¹H NMR (400 MHz, CDCl₃): δ 5.68 (ddd, J= 17.3, 10.6, 6.5 Hz, 1H), 4.88-5.02 (m, 2H), 4.24-4.32 (m, 2H), 3.93-4.03 (m, 1H), 2.52-2.63 (m, 1H), 1.95 (ddd, J= 12.3, 9.2, 7.3 Hz, 1H), 1.57-1.69 (m, 1H), 1.40 (s, 3H), 1.22 (s, 3H), 0.82 (s, 9H), 0.00 ppm (s, 6H). GC-MS m/z: [M-TBDMSi] Found 183.0, rt = 5.55 min. The spectroscopic results are consistent with reported literature data.[25]

Synthesisof((3aR,4R,6S,6aR)-6-((tert-butyldimethylsilyl)oxy)-2,2-dimethyltetra-hydro-4H-cyclopenta[d][1,3]dioxol-4-yl)methanol14.tert-

Butyl(((3aR,4S,6R,6aR)-2,2-dimethyl-6-vinyltetrahydro-4H-

cyclopenta[*d*][1,3]dioxol-4-yl)oxy)dimethylsilane **13** (27.5 g, 92.1 mmol, 1.00 eq) was dissolved in CH₂Cl₂ (800 ml) and the mixture was cooled to -78°C. Ozone was generated from oxygen gas with an ozone generator (Fischer OZ500/5 apparatus) and bubbled through the cooled solution via a glass pipet. A blue color was observed after 1.5 hours and ozone was added for an additional 20 minutes at -78°C. Subsequently, the mixture was flushed with nitrogen for 5 minutes until a colorless solution was obtained and dimethyl sulfide (40.6 ml, 553 mmol, 6.00 eq) was added at -78°C. The flow of nitrogen gas was stopped, and the mixture was stirred for 1 hour while the temperature gradually increased to -40°C. The mixture was concentrated *in vacuo* at 30°C to a minimal volume and the resulting yellow oil was dissolved in methanol (460 ml) and water (230 ml). The solution was cooled to 0°C and sodium

borohydride (31.3 g, 829 mmol, 9.00 eq) was added portion wise. The ice bath was removed after 1.5 hours and stirring was continued at room temperature. After 4 hours, the mixture was diluted in CH₂Cl₂ (500 ml) and NH₄Cl (ag. sat. 150 ml) was added. The product was extracted in CH₂Cl₂ (3x 500 ml) and combined organic layers were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo to afford the title compound 14. An analytical sample was purified by column chromatography over silica gel for characterization (gradient elution: n-heptane/EtOAc from 1:0 to 0:1). Colorless oil. Yield: 94% (26.3 g). ¹H NMR (400 MHz, CDCl₃): δ 4.37-4.42 (m, 2H), 4.10-4.15 (m, 1H), 3.55-3.62 (m, 1H), 3.45-3.53 (m, 1H), 2.16-2.26 (m, 1H), 2.01 (dt, J= 12.7, 8.2 Hz, 1H), 1.86 (br s, 1H), 1.57-1.66 (m, 1H), 1.49 (s, 3H), 1.31 (s, 3H), 0.91 (s, 9H), 0.09 ppm (d, J= 2.9 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃): δ 111.7, 82.0, 80.9, 72.7, 64.3, 44.7, 34.4, 26.5, 26.0, 24.9, 18.4, -4.5 ppm. GC-MS m/z: [M-TBDMSi] Found 187.1, rt = 6.55 min. The spectroscopic results are consistent with reported literature data.^[11]

Synthesis of tert-butyl(((3aR,4S,6aR)-2,2-dimethyl-6-methylenetetrahydro-4H-cyclopenta[d][1,3]dioxol-4-

yl)oxy)dimethylsilane 15. ((3a*R*,4*R*,6*S*,6a*R*)-6-((*tert*-Butyldimethylsilyl)oxy)-2,2-dimethyltetra-hydro-4*H*-

cyclopenta[d][1,3]dioxol-4-yl)methanol 14 (26.3 g, 86.9 mmol, 1.00 eq) was dissolved in THF (450 ml). Imidazole (14.8 g, 217 mmol, 2.50 eq) and triphenylphosphine (26.4 g, 95.6 mmol, 1.10 eq) were added followed by the portion wise addition of iodine (27.9 g, 108.7 mmol, 1.25 eq) at room temperature over 20 minutes. After 30 minutes after complete addition, the mixture was concentrated to a minimal volume in vacuo and n-heptane (500 ml) was added. Triphenylphosphine oxides were precipitated and the mixture was sonicated for 30 minutes. The solids were separated by filtration and rinsed with *n*-heptane (100 ml). To the filtrate was added sodium thiosulfite (aq. sat. 150 ml) and the product was extracted in n-heptane (3x 400 ml). Combined organic fractions were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue (30.6 g) was dissolved in THF (500 ml) and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 11.9 ml, 79.3 mmol, 1.10 eq) was added. The mixture was heated to 65°C for 1.5 hours, after which an additional portion of DBU (2.16 ml, 14.4 mmol, 0.20 eq) was added and heating was continued for 1.5 hours. Precipitated salts were removed via filtration at room temperature and the solids were rinsed with THF. The filtrate was concentrated to a minimal volume in vacuo and n-heptane (500 ml) and brine (150 ml) were added. The organic layer was washed with brine (3x 150 ml), dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: nheptane/EtOAc from 1:0 to 1:1). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 15. Colorless liquid. Yield: 67% (5 steps from 12, 18.3 g, 64.5 mmol). ¹H NMR (400 MHz, CDCl₃): δ 5.14-5.20 (m, 1H), 5.11 (dd, J= 2.6, 1.0 Hz, 1H), 4.62 (d, J= 5.7 Hz, 1H), 4.46 (t, J= 5.1 Hz, 1H), 3.90 (ddd, J= 11.2, 6.5, 4.7 Hz, 1H), 2.67 (ddtd, J= 13.9, 11.1, 2.7, 1.2 Hz, 1H), 2.29-2.35 (m, 1H), 1.50 (s, 3H), 1.35 (s, 3H), 0.92 (s, 9H), 0.11 ppm (d, J= 2.8 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃): δ 145.6, 113.6, 111.2, 80.8, 80.4, 72.3, 37.5, 26.4, 26.0, 24.7, 18.4, -4.5, -4.6 ppm. GC-MS m/z: [M-TBDMSi] Found 169.1, rt = 5.22 min.

Synthesis of (1*R*,3a'*R*,6'*S*,6a'*R*)-6'-((*tert*-Butyldimethylsilyl)oxy)-2,2-dichloro-2',2'dimethyltetrahydrospiro[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-3-one 16. Activation of Zinc:^[26] Zinc powder (25.0 g, 0.380 mol) was added to a two-necked round bottomed flask (500 ml) containing demineralized water (100 ml) and the solution was degassed with nitrogen for 15 minutes. Subsequently, copper(II) sulfate (1.85 g, 11.5 mmol) was added and the stirring solution was degassed for 45 minutes. The mixture was filtered, and the solids were washed with degassed water (250 ml) and degassed acetone (250 ml), respectively. The zinc-copper couple was dried *in vacuo* for 12 hours. *[2+2]-cycloaddition: tert*-Butyl(((3aR,4S,6aR)-2,2-dimethyl-6-methylenetetrahydro-4H-cyclopenta[d][1,3]dioxol-4-

yl)oxy)dimethylsilane 15 (2.50 g, 8.79 mmol, 1.00 eq) was dissolved in anhydrous diethyl ether (70 ml) dried over molecular sieves and Zn(Cu) (7.93 g, 61.5 mmol, 7.00 eq) was added. Trichloroacetyl chloride (2.45 ml, 22.0 mmol, 2.50 eq), dissolved in anhydrous diethyl ether (20 ml) was charged in a glass syringe and added dropwise (6.5 ml/hour) at room temperature over a period of 3 hours. The mixture was filtered over Celite to remove the solids and rinsed with diethyl ether (400 ml). The organic layer was washed with NaHCO₃ (aq. sat. 3x 200 ml) and brine (2x 150 ml), dried (MgSO₄), filtered and the filtrate was concentrated in vacuo to afford the title compound 16. Yellow oil. Yield: 97% (3.36 g, intermediate). ¹H NMR (400 MHz, CDCI₃): δ 4.79 (dd, *J*= 5.8, 1.0 Hz, 1H), 4.55 (t, J= 5.3 Hz, 1H), 4.11 (dt, J= 9.8, 5.1 Hz, 1H), 3.65 (d_{AB}, J= 18.3 Hz, 1H), 3.12 (d_{AB}, J= 18.3 Hz, 1H), 2.36 (dd, J= 12.9, 5.4 Hz, 1H), 2.15 (dd, J= 12.9, 9.8 Hz, 1H), 1.48 (s, 3H), 1.37 (s, 3H), 0.91-0.93 (m, 9H), 0.12 ppm (d, J= 2.2 Hz, 6H). ¹³C NMR (CDCl₃, 101MHz): δ 191.8, 112.3, 90.5, 80.7, 80.1, 71.5, 51.7, 49.2, 39.6, 26.1, 25.9, 24.7 18.4, -4.6, -4.9 ppm.

Synthesis of (3a'R,6'S,6a'R)-6'-((tert-Butyldimethylsilyl)oxy)-2',2'-dimethyltetrahydrospiro-[cyclobutane-1,4'-cyclopenta[d][1,3]dioxol]-3-one 17.

(1*R*,3a'*R*,6'*S*,6a'*R*)-6'-((*tert*-Butyldimethylsilyl)oxy)-2,2-dichloro-2',2'-dimethyltetrahydrospiro[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-3-one 16 (3.36 g, 8.50 mmol, 1.00 eq) was dissolved in THF (80 ml) and zinc (5.56 g, 85.0 mmol, 10.0 eq) followed by glacial acetic acid (3.89 ml, 68.0 mmol, 8.00 eq) were added. The mixture was heated to 70°C for 5 hours and filtered over Celite, rinsed with THF (100 m) and concentrated to a minimal volume in vacuo. The residue was dissolved in EtOAc (300 ml) and washed with NaHCO₃ (3x 100 ml) and brine (3x 100ml). The organic layer was dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified via silica gel chromatography (gradient elution: n-heptane/EtOAc: 1/0 to 2/3). Fractions containing the product were combined and the solvent was removed in vacuo to afford the title compound 17. Colorless oil. Yield: 67% (2 steps from 15, 1.90 g, 5.82 mmol). ¹H NMR (400 MHz, CDCl₃): δ 4.49 (t, J= 5.1 Hz, 1H), 4.29-4.34 (m, 1H), 3.88 (dt, J= 10.9, 5.3 Hz, 1H), 3.36 (ddd, J= 18.3, 4.1, 2.4 Hz, 1H), 2.89-2.99 (m, 1H), 2.79-2.87 (m, 1H), 2.68-2.75 (m, 1H), 2.19 (t, J= 11.4 Hz, 1H), 1.82 (dd, J= 11.8, 5.7 Hz, 1H), 1.58 (s, 1H), 1.48 (s, 3H), 1.34 (s, 3H), 0.90-0.94 (m, 9H), 0.11 ppm (d, J= 2.8 Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3): δ 206.0, 111.1, 85.2, 80.2, 72.3, 56.7, 52.8, 41.3, 34.4, 26.0, 26.0, 24.5, 18.4, -4.4, -4.7 ppm. GC-MS m/z: [M-TBDMSi] Found 211.1, rt = 6.91 min.

General procedure 1: Enzymatic cyclobutanone reduction. Synthesis of (2*S*,4*r*,6*R*,7*R*,8*S*)-2-hydroxy-6methoxy-5-oxaspiro[3.4]octane-7,8-diyl bis(2,2dimethylpropanoate) 22b. The Codex® Ketoreductase (KRED) screening kit was purchased from Codexis®. Detailed protocols for the initial enzyme screening can be found in the

information. (6R,7R,8S)-6-Methoxy-2-oxo-5supplementary oxaspiro[3.4]octane-7,8-diyl bis(2,2-dimethylpropanoate) (1.00 g, 2.81 mmol, 1.00 eq) was dissolved in *i*PrOH (2.1 ml) in a falcon tube. The crude enzyme (B02, 50 mg) was added to a wellmixed solution of H₂O (Milli-Q, 12.6 ml) and KRED Recycle Mix P buffer (100 mg) [Na₃PO₄ (128 mM), MgSO₄ (1.7 mM), NADP⁺ (1.1 mM) at pH 7.0]. The aqueous solution was gently shaken to avoid foam formation until a homogeneous solution was obtained and was subsequently added all at once to ketone 20 dissolved in iPrOH. The mixture was shaken at 33°C for 24 hours. To the falcon tube was added EtOAc (10 ml) and the mixture was mixed vigorously and centrifuged (4000 rpm, 2 min). The organic layer was collected and this extraction with EtOAc was repeated twice. Combined organic fractions were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: nheptane/EtOAc from 4:1 to 0:1). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 22b. White solid. Yield: 92% (910 mg, 2.59 mmol). ¹H NMR (400 MHz, CDCl₃): δ 5.30 (d, J= 4.5 Hz, 1H), 5.13-5.17 (m, 1H), 4.84 (d, J= 1.6 Hz, 1H), 4.46-4.60 (m, 1H), 3.39 (s, 3H), 2.54-2.63 (m, 1H), 2.39-2.50 (m, 2H), 2.20-2.33 (m, 2H), 1.25 (s, 9H), 1.20 ppm (s, 9H). ¹³C NMR (101 MHz, CDCl₃): δ 177.6, 177.0, 105.5, 80.6, 75.2, 74.8, 62.6, 55.3, 44.3, 41.6, 39.0, 38.8, 27.2, 27.1 ppm. LC-MS (method 3, ESI+) m/z: [M+H]+ Found 359.3, rt = 2.06 min.

(1s,3R,3a'R,6'S,6a'R)-6'-((*tert*-Butyldimethylsilyl)oxy)-2',2'dimethyltetrahydrospiro-[cyclobutane-1,4'-

cyclopenta[*d*][1,3]dioxol]-3-ol (**18a**). The reaction was performed according to general procedure 1, by employing KRED enzyme D03 (65 mg), KRED *recycle mix P* buffer (150 mg), Milli-Q H₂O (9 ml) and *i*PrOH (1.45 ml). Colorless viscous oil. Yield: 95% (240 mg, 0.73 mmol). ¹H NMR (400 MHz, CDCl₃): δ 4.32 (t, *J*= 5.17 Hz, 1 H), 4.22 - 4.30 (m, 1 H), 4.18 (d, *J*= 5.28 Hz, 1 H), 3.77 - 3.85 (m, 1 H), 2.31 (br s, 1 H), 2.02 - 2.25 (m, 3 H), 1.85 - 1.93 (m, 1 H), 1.62 - 1.72 (m, 2 H), 1.42 (s, 3 H), 1.29 (s, 3 H), 0.90 (s, 9 H), 0.08 ppm (d, *J*= 1.76 Hz, 6 H). ¹³C NMR (101 MHz, CDCl₃): δ 110.42, 87.35, 79.59, 72.09, 63.86, 43.13, 40.97, 39.08, 35.41, 25.98, 24.48, 18.41, -4.4, -4.6 ppm. GC-MS m/z: [M-TBDMSi] Found 213.1, rt = 6.07 min.

(1r,3S,3a'R,6'S,6a'R)-6'-((tert-butyldimethylsilyl)oxy)-2',2'dimethyltetrahydrospiro-[cyclobutane-1,4'-

cyclopenta[*d*][*1*,3]*dioxo*[*1*-3-*ol* (*18b*). The reaction was performed according to general procedure 1, by employing KRED enzyme H07 (155 mg), KRED *recycle mix P* buffer (150 mg), Milli-Q H₂O (9 ml) and *i*PrOH (1.45 ml). Colorless viscous oil. Yield: 51% (129 mg, 0.39 mmol, 41% of **17** recovered). ¹H NMR (400 MHz, CDCl₃): δ 4.37 (t, *J*= 5.17 Hz, 1 H), 4.28 (quin, *J*= 7.32 Hz, 1 H), 4.16 (d, *J*= 5.28 Hz, 1 H), 3.76 (dt, *J*= 11.17, 5.53 Hz, 1 H), 2.59 (ddd, *J*= 11.99, 6.93, 5.28 Hz, 1 H), 2.22 (br s, 1 H), 2.13 (ddd, *J*= 11.99, 7.59 Hz, 1 H), 1.93 (t, *J*= 11.44 Hz, 1 H), 1.84 (dd, *J*= 11.99, 7.59 Hz, 1 H), 1.59 - 1.71 (m, 2 H), 1.43 (s, 3 H), 1.30 (s, 3 H), 0.89 (s, 9 H), 0.07 ppm (d, *J*= 1.98 Hz, 6 H). ¹³C NMR (101 MHz, CDCl₃): δ 110.50, 84.90, 79.94, 71.62, 63.20, 43.28, 43.01, 37.64, 35.55, 25.98, 25.97, 24.49, 18.39, -4.4, -4.7 ppm.

Synthesis of *tert*-butyl(((3a'*R*,6'S,6a'*R*)-2',2'-dimethyl-3-methylenetetrahydrospiro-[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-6'-yl)oxy)dimethylsilane26.Methyltriphenylphosphonium bromide (1.45 g, 3.98 mmol, 1.30eq) was weighed in an oven dried vial and THF (12.0 ml) wasadded. The heterogeneous solution was cooled to 0°C and

potassium *tert*-butoxide (3.98 ml, 1M in THF, 3.98 mmol, 1.30 eq) was added dropwise. The mixture was stirred at 0°C for 20 minutes. The freshly prepared Wittig reagent was added dropwise via syringe to (3a'R,6'S,6a'R)-6'-((tert-butyldimethylsilyl)oxy)-2',2'- dimethyltetrahydrospiro-[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-3-one 17 (1.00 g, 3.06 mmol, 1.00 eq) dissolved in THF (12.0 ml) at 0°C. The yellow mixture was stirred for 1.5 hours at 0°C followed by 1.5 hours stirring at room temperature. The mixture was concentrated to a minimal volume vacuo and redissolved in *n*-heptane (300 ml). in Triphenylphosphine oxide was precipitated and the mixture was sonicated for 5 minutes, filtered and the filtrate was washed with NH₄Cl (aq. sat. 2x 50 ml) and brine (2x 50 ml). The organic layer was dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: n-heptane/EtOAc from 1:0 to 7:3). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 26. Colorless oil. Yield: 94% (931 mg, 2.88 mmol). ¹H NMR (400 MHz, CDCl₃): δ 4.81 (quin, J= 2.4 Hz, 1H), 4.78 (quin, J= 2.4 Hz, 1H), 4.40 (t, J= 5.1 Hz, 1H), 4.26 (dd, J= 5.5, 0.9 Hz, 1H), 3.80 (dt, J= 11.2, 5.5 Hz, 1H), 2.87 (dd, J= 16.1, 2.2 Hz, 1H), 2.54 (dq, J= 15.9, 2.4 Hz, 1H), 2.31-2.46 (m, 2H), 1.95 (t, J= 11.4 Hz, 1H), 1.76 (dd, J= 11.7, 5.7 Hz, 1H), 1.45 (s, 3H), 1.32 (s, 3H), 0.91 (s, 9H), 0.10 ppm (d, J= 2.2 Hz, 6H). ¹³C NMR (101 MHz, CDCl₃): δ 144.7, 110.4, 106.8, 85.3, 79.9, 72.0, 42.5, 41.2, 39.2, 36.8, 26.0, 26.0, 24.5, 18.4, -4.4, -4.6 ppm.

Synthesis of (6*R*,7*R*,8*S*)-6-methoxy-2-methylene-5oxaspiro[3.4]octane-7,8-diyl bis(2,2-dimethylpropanoate) 28. (6*R*,7*R*,8*S*)-6-Methoxy-2-oxo-5-oxaspiro[3.4]octane-7,8-diyl

bis(2,2-dimethylpropanoate) 20 (2.00 g, 5.50 mmol, 1.00 eq) was weighed in a three neck 100 ml flask equipped with a reflux condenser, thermometer and CaCl2 tube. To this flask was added a solution of bis(cyclopentadienyl)dimethyltitanium (32, 39.4 ml, 5 wt% in toluene, 7.97 mmol, 1.45 eq, CAS: 1271-66-5). The flask was covered from light with aluminium foil and heated to 70°C. The reaction was stirred for 17 hours after which full conversion was observed. The mixture was concentrated to a minimal volume in vacuo and to the residue was added n-heptane (100 ml). The solids were sonicated for 5 minutes and removed via filtration over celite (rinsed with n-heptane). The organic layer was concentrated to a minimal volume in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: n-heptane/EtOAc from 1:0 to 3:7). Fractions containing the product were collected and the solvent was evaporated to afford the title compound 28. Colorless oil. Yield: 58% (1.14 g, 3.19 mmol). ¹H NMR (500 MHz, CDCI₃): δ 5.42 (d, J= 4.5 Hz, 1H), 5.17, (dd, J= 4.5, 1.7 Hz, 1H), 4.85-4.83 (m, 3H), 3.39 (s, 3H), 3.09-3.05 (m, 2H), 2.94-2.85 (m, 2H), 1.21 (s, 9H), 1.20 ppm (s, 9H). ¹³C NMR (125 MHz, CDCl₃): δ 177.4, 177.2, 139.7, 107.6, 105.7, 80.6, 75.6, 74.5, 55.6, 44.8, 41.5, 39.2, 39.0, 27.4, 27.3 ppm. LC-MS (method 1, ESI+) m/z: [M+Na]+ Found 376.86.

Synthesis of (3a'*R*,6'S,6a'*R*)-6'-((*tert*-butyldimethylsilyl)oxy)-2',2'-dimethyl-3-

phenyltetrahydrospiro[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-3-ol 25. (3a'R,6'S,6a'R)-6'-((*tert*-Butyldimethylsilyl)oxy)-2',2'-dimethyltetrahydrospiro[cyclobutane-1,4'-cyclopenta[d][1,3]dioxol]-3-one **17** (350 mg, 1.07 mmol, 1.00 eq) was dissolved in anhydrous THF (5.50 ml) and cooled to -20°C. Subsequently, phenylmagnesium bromide (0.44 ml, 2.9 M in THF, 1.29 mmol, 1.45 eq) was added dropwise and the mixture

was stirred at -10°C for 1 hour. The mixture was quenched at -10°C by adding NH₄CI (aq. sat. 20 ml) and the product was extracted in CH₂Cl₂ (1x 70 ml, 2x 50 ml). Combined organic layers were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: n-heptane/EtOAc from 1:0 to 2:3). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 25 as a 1:1 mixture of diastereoisomers. An analytical sample was purified via column chromatography for characterization of 25a and 25b. Colorless oil. Yield: 96% (417 mg, 1.03 mmol, combined). 25a: ¹H NMR (400 MHz, CDCl₃): δ 7.31-7.35 (m, 2H), 7.22-7.29 (m, 2H), 7.12-7.19 (m, 1H), 4.22 (t, J= 5.1 Hz, 1H), 3.96 (d, J= 5.3 Hz, 1H), 3.69-3.82 (m, 1H), 2.87 (dd, J= 13.2, 1.4 Hz, 1H), 2.23 (dd, J= 12.2, 2.0 Hz, 1H), 2.15 (dd, J= 13.0, 2.8 Hz, 1H), 2.06 (dd, J= 12.2, 2.8 Hz, 1H), 1.94-2.01 (m, 2H), 1.34 (s, 3H), 1.14 (s, 3H), 0.81 (s, 9H), 0.00 ppm (s, 6H). ¹³C NMR (101 MHz, CDCl3): δ 146.6, 128.5, 127.4, 125.0, 110.4, 85.9, 79.8, 73.7, 71.8, 46.9, 42.7, 40.7, 35.8, 26.1, 26.0, 24.5, 18.5, -4.4, -4.5 ppm. LC-MS (method 2, ESI+) m/z: [M+H]⁺ Found 405.4 (Rt: 1.37 min, poor mass response). 25b: ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.46 (m, 4H), 7.26-7.32 (m, 1H), 4.61 (d, J= 5.5 Hz, 1H), 4.44 (t, J= 5.2 Hz, 1H), 3.81 (dt, J= 10.7, 5.4 Hz, 1H), 2.62-2.69 (m, 1H), 2.53-2.61 (m, 1H), 2.27-2.36 (m, 1H), 2.18-2.27 (m, 1H), 2.13 (s, 1H), 1.94 (t, J= 11.4 Hz, 1H), 1.59 (dd, J= 11.7, 5.5 Hz, 1H), 1.48 (s, 3H), 1.36 (s, 3H), 0.90 (s, 9H), 0.08 ppm (d, J= 4.4 Hz, 6H). ¹³C NMR (101 MHz, CDCI₃): δ 146.6, 128.5, 127.4, 125.0, 110.7, 86.5, 80.0, 74.4, 71.8, 46.6, 42.1, 40.9, 36.6, 26.1, 26.0, 24.6, 18.5, -4.4, -4.6 ppm. LC-MS (method 2, ESI+) m/z: [M+H]+ Found 405.4 (Rt: 1.39 min, poor mass response).

General procedure 2: Wittig reaction. Synthesis of 2-((3a'S,6'*R*,6a'*R*)-6'-methoxy-2',2'-dimethyldihydro-6'*H*-spiro [cyclobutane-1,4'-furo[3,4-*d*][1,3]dioxol]-3-

ylidene)acetonitrile **30**. (3a'S,6'R,6a'R)-6'-Methoxy-2',2'dimethyldihydro-6'H-spiro[cyclobutane-1,4'-furo[3,4-

d][1,3]dioxol]-3-one 19 (100 mg, 0.44 mmol, 1.00 eq) was dissolved in THF (1.5 ml) and cyanomethyltriphenylphosphonium ylide 29 (191 mg, 0.66 mmol, 1.50 eq) was added. The mixture was heated to 65°C for 21 hours. The mixture was concentrated to a minimal volume in vacuo and the residue was purified by column chromatography over silica gel (gradient elution: nheptane/EtOAc from 99:1 to 1:9). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 30 as a 1:1 mixture of diastereoisomers. An analytical sample was purified by silica gel chromatography for the characterization of both diastereoisomers 30a and 30b. Colorless oil. Yield: 55% (60 mg, 0.24 mmol, combined). 30a: 1H NMR (500 MHz, CDCl₃): δ 5.28-5.27 (m, 1H), 4.88 (s, 1H), 4.61 (d, J= 5.8 Hz, 1H), 4.56 (d, J= 5.8 Hz, 1H), 3.34 (s, 3H), 3.34-3.26 (m, 4H), 3.13-3.10 (m, 1H), 3.01-3.00 (m, 2H), 1.39 (s, 3H), 1.30 ppm (s, 3H). ¹³C NMR (125 MHz, CDCl₃): δ 162.6, 116.0, 112.9, 108.4, 93.9, 85.2, 83.9, 81.7, 55.2, 46.3, 40.3, 26.5, 25.3 ppm. **30b**: ¹H NMR (500 MHz, CDCl₃): δ 5.26-5.24 (m, 1H), 4.89 (s, 1H), 4.60 (d, J= 5.8 Hz, 1H), 4.55 (d, J= 5.8 Hz, 1H), 3.48-3.42 (m, 1H), 3.34 (s, 3H), 3.22-3.17 (m, 1H), 2.97-2.93 (m, 1H), 2.89-2.84 (m, 1H), 1.41 (s, 3H), 1.31 ppm (s, 1H). ¹³C NMR (125 MHz, CDCI₃): δ 162.6, 116.0, 113.0, 108.5, 93.9, 85.2, 83.9, 81.8, 55.3, 46.5, 40.2, 26.5, 25.4 ppm.

(6R,7R,8S)-2-(Cyanomethylene)-6-methoxy-5-

oxaspiro[3.4]octane-7,8-diyl bis(2,2-dimethylpropanoate) (**31**). The reaction was performed according to general procedure 2.

Colorless oil. Yield: 42% (22 mg, 0.059 mmol, combined). ¹H NMR (500 MHz, CDCl₃): δ 5.42 (d, *J*= 4.46 Hz, 1H), 5.41 (d, *J*= 4.46 Hz, 1H), 5.24-5.22 (m, 2H), 5.19-5.17 (m, 2H), 4.84 (s, 1H), 4.82 (s, 1H), 3.43-3.33 (m, 7H), 3.32- 3.24 (m, 4H), 3.21-3.16 (m, 1H), 3.10-2.98 (m, 4H), 1.21 (s, 18H), 1.20 ppm (s, 18H). ¹³C NMR (125 MHz, CDCl₃): δ 177.5, 177.2, 177.1, 177.0, 161.9, 161.8, 116.0, 115.8, 105.8, 105.7, 93.5, 93.3, 79.7, 79.6, 75.1, 74.2, 74.0, 55.7, 55.6, 45.5, 45.1, 42.7, 42.6, 39.2, 39.1, 39.1, 39.0, 27.3 ppm.

Synthesis of 7-(((1s,3R,3a'R,6'S,6a'R)-6'-((*tert*butyldimethylsilyl)oxy)-2',2'-dimethyltetrahydro-spiro-[cyclobutane-1,4'-cyclopenta[d][1,3]dioxol]-3-

(1r,3S,3a'R,6'S,6a'R)-6'-((tertyl)oxy)quinoline 33. Butyldimethylsilyl)oxy)-2',2'-dimethyltetrahydrospiro[cyclobutane-1,4'-cyclopenta[d][1,3]dioxol]-3-ol 18b (111 mg, 0.34 mmol, 1.00 eq) was dissolved in THF (3 ml) and triphenylphosphine (106 mg, 0.41 mmol, 1.20 eq), 7-hydroxyquinoline (58.8 mg, 0.41 mmol, 1.20 eq) and diisopropyl azodicarboxylate (0.08 ml, 0.41 mmol, 1.2 eq) were added at room temperature. The mixture was stirred for 17 hours at room temperature and additional portions of triphenylphosphine (53.0 mg, 0.21 mmol, 0.6 eq), 7hydroxyquinoline (29.4 mg, 0.21 mmol, 0.6 eq) and diisopropyl azodicarboxylate (0.04 ml, 0.21 mmol, 0.6 eq) were added. After 7 hours, the mixture was concentrated in vacuo and the residue was purified via silica gel chromatography (gradient: nheptane:EtOAc 1:1 to 1:4). Fractions containing the product were combined and the solvent was removed in vacuo to afford the title compound 33. Colorless oil. Yield: 80% (123 mg, 0.27 mmol). ¹H NMR (400 MHz, CDCl₃): δ 8.68 (dd, *J*= 4.4, 1.8 Hz, 1H), 7.93 (dd, J= 8.3, 1.4 Hz, 1H), 7.56 (d, J= 9.0 Hz, 1H), 7.12-7.15 (m, 1H), 7.11 (s, 1H), 7.07-7.14 (m, 1H), 7.04 (dd, J= 8.8, 2.4 Hz, 1H), 4.72 (quin, J= 6.8 Hz, 1H), 4.25 (t, J= 5.1 Hz, 1H), 4.14 (d, J= 5.3 Hz, 1H), 3.76 (dt, J= 10.9, 5.6 Hz, 1H), 2.31-2.44 (m, 2H), 2.17-2.31 (m, 1H), 1.85-1.94 (m, 2H), 1.70 (dd, J= 11.8, 5.6 Hz, 2H), 1.31 (s, 3H), 1.18 (s, 3H), 0.81 (s, 9H), 0.00 ppm (s, 6H). ¹³C NMR (101 MHz, CDCl₃): δ 158.4, 150.5, 149.9, 135.7, 128.9, 123.5, 120.2, 119.0, 110.6, 108.8, 87.1, 79.8, 72.1, 68.8, 41.5, 40.7, 37.0, 35.9, 26.1, 24.6, 18.5, -4.4, -4.5 ppm. LC-MS (method 3, ESI+) m/z: [M+H]⁺ Found 456.3 (Rt: 2.76 min).

Synthesis of 7-(((1*s*,3*R*,3a'*R*,6'*R*,6a'*S*)-6'-(4-chloro-7*H*-pyrrolo[2,3-*d*]pyrimidin-7-yl)-2',2'-

dimethyltetrahydrospiro[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-3-yl)oxy)-quinoline 36. 7-(((1*s*,3*R*,3a'*R*,6'*S*,6a'*R*)-6'-((*tert*-Butyldimethylsilyl)oxy)-2',2'-

dimethyltetrahydro-spiro-[cyclobutane-1,4'-

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cyclopenta[d][1,3]dioxol]-3-yl)oxy)quinoline 33 (123 mg, 0.27
mmol, 1.00 eq) was dissolved in THF (3.50 ml) and tert-
butylammonium fluoride (0.54 ml, 1 M in THF, 2 eq) was added.
The mixture was stirred for 1 hour at room temperature and
concentrated to a minimal volume in vacuo. The residue was
dissolved in EtOAc (50 ml) and washed with NaHCO<sub>3</sub> (1x 25 ml)
and brine (3x 25ml). The organic layer was dried (MgSO<sub>4</sub>), filtered
and the filtrate was concentrated to a minimal volume in vacuo.
The residue was dissolved in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (2.00 ml) and
anhydrous pyridine (0.07 ml, 0.84 mmol, 3.00 eq) was added. The
solution was cooled to 0°C and trifluoromethanesulfonic
anhydride (0.07 ml, 0.39 mmol, 1.40 eq) was added dropwise.
The mixture was stirred for 30 minutes at 0°C and then diluted in
CH<sub>2</sub>Cl<sub>2</sub> (25 ml) and NaHCO<sub>3</sub> (aq. 5 ml) was added. The product
was extracted in CH<sub>2</sub>Cl<sub>2</sub> (3x 25 ml) and combined organic layers
were dried (MgSO<sub>4</sub>), filtered and the filtrate concentrated in vacuo.
The orange residue was used immediately further. 4-Chloro-7H-
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pyrrolo[2,3-d]pyrimidine (35b, 206 mg, 1.07 mmol, 4.00 eq) was dissolved in anhydrous DMF (2.0 ml) and the crude triflate residue dissolved in DMF (1.5 ml) was added dropwise at -10°C during 15 minutes. The mixture was stirred for 1.5 hours at -15°C and then 30 minutes at room temperature. The reaction was guenched with NH₄Cl (aq. sat. 50 ml) and the product was extracted with CH₂Cl₂ (3x 80 ml). Combined organic layers were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. A mixture of the desired substitution product 36 (75%) and elimination side product 37 (25%) was obtained, as evidenced via NMR analysis. The residue was purified by column chromatography over silica gel (gradient elution: n-heptane/EtOAc from 1:0 to 0:1). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 36. White solid. Yield: 47% (3 steps, 59 mg, 0.12 mmol). ¹H NMR (400 MHz, CDCl₃): δ 8.81 (dd, J= 4.3, 1.8 Hz, 1H), 8.66 (s, 1H), 8.06 (dd, J= 8.1, 1.2 Hz, 1H), 7.69 (d, J= 8.5 Hz, 1H), 7.15-7.36 (m, 5H), 6.64 (d, J= 3.3 Hz, 1H), 5.10 (dd, J= 6.7, 3.9 Hz, 1H), 4.89-5.01 (m, 1H), 4.69-4.85 (m, 2H), 2.78 (dd, J= 12.0, 7.1 Hz, 1H), 2.34-2.63 (m, 6H), 1.86 (br s, 1H), 1.52 (s, 3H), 1.31 ppm (s, 3H). ¹³C NMR (101 MHz, CDCl₃): δ 158.3, 152.4, 150.9, 150.6, 150.6, 149.8, 135.7, 129.0, 127.8, 123.6, 120.1, 119.1, 118.1, 113.2, 108.7, 100.0, 85.7, 83.9, 67.9, 61.5, 42.8, 40.3, 39.8, 36.6, 26.6, 24.8 ppm. HRMS (ESI+) m/z: [M+H]⁺ Calcd. for C₂₆H₂₆ClN₄O₃ 477.1693; Found 477.1691 (Rt: 10.96 min). 7-(((1s,3S,3a'R,6a'S)-2',2'-Dimethyl-3a',6a'dihydrospiro[cyclobutane-1,4'-cyclopenta-[d][1,3]dioxol]-3-

yl)oxy)quinoline **37**: Colorless oil. Yield: **10%** (3 steps, 9 mg, 0.03 mmol), colorless oil. ¹H NMR (400 MHz, CDCl₃): δ 8.81 (dd, *J*=4.3, 1.8 Hz, 1H), 8.06 (dd, *J*=8.1, 1.6 Hz, 1H), 7.69 (d, *J*=9.0 Hz, 1H), 7.22-7.30 (m, 2H), 7.18 (dd, *J*=8.7, 2.6 Hz, 1H), 5.97 (d, *J*=5.7 Hz, 1H), 5.79 (dd, *J*=5.7, 1.6 Hz, 1H), 5.13 (dd, *J*=5.7, 1.2 Hz, 1H), 4.92 (t, *J*=6.9 Hz, 1H), 4.51 (d, *J*=6.1 Hz, 1H), 2.77 (dd, *J*=12.2, 6.9 Hz, 1H), 2.49-2.62 (m, 2H), 2.41 (dd, *J*=11.8, 6.9 Hz, 1H), 1.36 (s, 3H), 1.35 ppm (s, 3H). ¹³C NMR (101 MHz, CDCl₃): δ 158.3, 150.5, 149.8, 140.4, 135.7, 129.5, 128.9, 123.5, 120.2, 119.0, 111.1, 108.7, 86.4, 84.6, 68.0, 46.9, 40.7, 36.7, 27.5, 26.2 ppm. LC-MS (method 2, ESI⁺) m/z: [M+H]⁺ Found 324.2 (Rt: 1.12 min).

Synthesis of 7-((1s,3R,3a'R,6'R,6a'S)-2',2'-dimethyl-3-(quinolin-7-yloxy)tetrahydrospiro[cyclobutane-1,4'cyclopenta[d][1,3]dioxol]-6'-yl)-7H-pyrrolo[2,3-d]pyrimidin-4amine 41. 7-(((1s,3R,3a'R,6'R,6a'S)-6'-(4-chloro-7H-pyrrolo[2,3d]pyrimidin-7-yl)-2',2'-dimethyltetrahydrospiro[cyclobutane-1,4'cyclopenta[d][1,3]dioxol]-3-yl)oxy)-quinoline 36 (46.0 mg, 0.10 mmol, 1.00 eq) was dissolved in 1,4-dioxane (10 ml) and NH_3 (28% in H_2O, 30 ml) was added. The mixture was heated to 90°C in a pressure reactor for 17 hours. The mixture was concentrated to a minimal volume in vacuo and CH2Cl2 (30 ml) and brine (15 ml) were added. The product was extracted in CH₂Cl₂ (3x 30 ml) and combined organic layers were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified via silica gel chromatography (gradient: CH₂Cl₂: MeOH from 99:1 to 85:15) and fractions containing the product were collected and the solvent was removed in vacuo to afford the title compound 41. Slight orange oil. Yield: 80% (35.5 mg, 0.08 mmol). ¹H NMR (400 MHz, CDCl₃) δ ppm 8.80 (dd, *J*= 4.29, 1.65 Hz, 1 H) 8.34 (s, 1 H) 8.05 (dd, J= 8.25, 1.21 Hz, 1 H) 7.68 (d, J= 9.02 Hz, 1 H) 7.21 -7.28 (m, 2 H) 7.16 (dd, J= 8.91, 2.53 Hz, 1 H) 6.90 (d, J= 3.74 Hz, 1 H) 6.37 (d, J= 3.52 Hz, 1 H) 5.36 (br s, 2 H) 5.09 (dd, J= 6.71, 3.41 Hz, 1 H) 4.91 (td, J= 7.26, 3.30 Hz, 1 H) 4.77 (quin, J= 7.04 Hz, 1 H) 4.68 (d, J= 6.60 Hz, 1 H) 2.75 (dd, J= 12.21, 7.15 Hz, 1 H) 2.29 - 2.52 (m, 5 H) 1.52 (s, 3 H) 1.33 ppm (s, 3 H). ¹³C NMR

 $\begin{array}{l} (101 \ \text{MHz}, \ \text{CDCl}_3): \ \delta = 158.4, \ 156.8, \ 151.8, \ 150.5, \ 150.5, \ 149.8, \\ 135.7, \ 128.9, \ 123.5, \ 123.3, \ 120.2, \ 119.0, \ 112.8, \ 108.8, \ 103.7, \\ 97.9, \ 86.0, \ 84.2, \ 68.1, \ 60.8, \ 42.9, \ 40.3, \ 39.9, \ 36.6, \ 26.5, \ 24.8 \ \text{ppm.} \\ \text{LC-MS} \ (\text{method}\ 2, \ \text{ESI}^+) \ m/z: \ [\text{M+H]}^+ \ \text{Found} \ 458.3 \ (\text{Rt} = 0.92 \ \text{min}). \end{array}$

Synthesis of (2R,4s,5R,6S,7R)-7-(4-amino-7H-

pyrrolo[2,3-*d*]pyrimidin-7-yl)-2-(quinolin-7-

yloxy)spiro[3.4]octane-5,6-diol 38. 7-((1s,3R,3a'R,6'R,6a'S)-2',2'-dimethyl-3-(quinolin-7-yloxy)tetrahydrospiro[cyclobutane-

1,4'-cyclopenta[d][1,3]dioxol]-6'-yl)-7H-pyrrolo[2,3-d]pyrimidin-4amine 41 (35 mg, 0.08 mmol, 1.00 eq) was dissolved in EtOH (1 ml) and HCI (0.77 ml, 1M in H₂O, 0.76 mmol, 10.0 eq) was added. The mixture was stirred at room temperature for 4 hours and HCI (0.77 ml, 1M in H₂O, 0.76 mmol, 10.0 eq) was added. After 3 hours, the mixture was diluted with water (8 ml) and lyophilized to afford the title compound 38. Slight orange oil. Yield: 82% (28.5 mg, 0.06 mmol, HCI salt). An analytical sample was purified via Prep SCF (Stationary phase: Chiralcel Diacel OJ 20 x 250 mm, Mobile phase: CO₂, EtOH + 0.4 *i*PrNH₂) to afford the title compound 38. Slight orange solid. Yield: 31% (10.0 mg, 0.025 mmol). ¹H NMR (400 MHz, DMSO-d₆): δ 8.80 (dd, J= 4.3, 1.7 Hz, 1H), 8.26 (dd, J= 8.1, 1.3 Hz, 1H), 8.06 (s, 1H), 7.88 (d, J= 8.8 Hz, 1H), 7.36 (dd, J= 8.1, 4.2 Hz, 1H), 7.21-7.30 (m, 3H), 6.91 (s, 2H), 6.57 (d, J= 3.5 Hz, 1H), 4.80-4.99 (m, 4H), 4.25 (q, J= 5.7 Hz, 1H), 3.82 (t, J= 5.0 Hz, 1H), 2.54-2.71 (m, 1H), 2.43-2.49 (m, 1H), 2.26-2.37 (m, 1H), 2.21 (dd, J= 11.2, 7.0 Hz, 1H), 2.06 ppm (dd, J= 13.1, 8.9 Hz, 1H). ¹³C NMR (101 MHz, DMSO-d₆): δ 157.9, 157.4, 151.3, 150.6, 149.8, 149.4, 135.6, 129.3, 123.0, 122.5, 119.5, 119.2, 108.6, 102.8, 98.6, 77.2, 75.3, 67.8, 59.8, 40.3, 39.6, 39.2, 36.2 ppm. HRMS (ESI+) m/z: [M+H]+ Calcd. for C23H24N5O3 418.1879; Found 418.1873 (Rt: 6.73 min).

Synthesis of (3a'*R*,6'S,6a'S)-2',2'-dimethyl-3methylenetetrahydrospiro[cyclobutane-1,4'-cyclopenta-

[d][1,3]dioxol]-6'-ol 42. *tert*-Butyl(((3a'R,6'S,6a'R)-2',2'-dimethyl-3-methylenetetrahydrospiro[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-6'-yl)oxy)dimethylsilane 26 (931 mg, 2.87 mmol, 1.00 eq) was dissolved in THF (2.00 ml) and tetrabutylammonium fluoride (10.0 ml, 1M in THF, 10.0 mmol, 3.50 eq) was added. The mixture was stirred at room temperature for 3 hours. The mixture was concentrated to a minimal volume in vacuo, dissolved in EtOAc (250 ml) and washed with NH₄Cl (aq. sat. 3x 50ml) and brine (3x 50 ml). The organic layer was dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: n-heptane/EtOAc from 1:0 to 0:1). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 42. Colorless oil. Yield: 92% (556 mg, 2.64 mmol). $^1\!H$ NMR (400 MHz, CDCl_3): δ 4.83 (quin, J= 2.3 Hz, 1H), 4.80 (quin, J= 2.4 Hz, 1H), 4.46-4.49 (m, 1H), 4.36-4.39 (m, 1H), 3.81 (br s, 1H), 2.82-2.88 (m, 1H), 2.60 (dq, J= 16.1, 2.4 Hz, 1H), 2.38-2.46 (m, 2H), 2.27-2.38 (m, 1H), 1.97 (dd, J= 12.0, 5.9 Hz, 1H), 1.74 (t, J= 11.4 Hz, 1H), 1.47 (s, 3H), 1.36 ppm (s, 3H). ¹³C NMR (101 MHz, CDCl₃): δ 144.1, 110.6, 107.2, 85.4, 78.7, 70.8, 41.7, 41.2, 39.5, 36.6, 25.9, 24.3 ppm.

Synthesis of 4-chloro-7-((3a'R,6'R,6a'S)-2',2'-dimethyl-3-methylenetetrahydrospiro-[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-6'-yl)-7H-pyrrolo[2,3-d]pyrimidine 43. (3a'R,6'S,6a'S)-2',2'-Dimethyl-3methylenetetrahydrospiro[cyclobutane-1,4'-cyclopenta-

[d][1,3]dioxol]-6'-ol **42** (647 mg, 3.08 mmol, 1.00 eq) was dissolved in anhydrous CH₂Cl₂ (20.0 ml) and pyridine (0.62 ml, 7.69 mmol, 2.50 eq) was added. The mixture was cooled to 0°C

and trifluoromethanesulfonic anhydride (0.57 ml, 3.39 mmol, 1.10 eq) was added dropwise. The mixture was stirred for 30 minutes at 0°C, diluted in CH₂Cl₂ (100 ml) and NaHCO₃ (aq. sat. 40 ml) was added. The product was extracted in CH₂Cl₂ (3x 100 ml) and combined organic layers were dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue was dissolved in anhydrous DMF (8 ml) and added dropwise over 15 min to a mixture of 35b (5.90 g, 30.8 mmol, 10.0 eq) in anhydrous DMF (35.0 ml) which was previously stirred for 30 minutes at 0°C. After addition, the mixture was stirred for 2 hours at 0°C and then warmed to room temperature and stirred for an additional 2 hours. The mixture was poured in NH₄Cl (aq. sat. 50 ml) and the product was extracted in EtOAc (3x 100 ml). Combined organic layers were washed with brine (3x 100 ml), dried (MgSO₄), filtered and the filtrate was concentrated in vacuo to a minimal volume. To the resulting powder, n-heptane (100 ml) was added and the mixture was sonicated for 10 minutes. The solids were filtered, rinsed with *n*-heptane and the filtrate was concentrated to a minimal volume in vacuo. The residue was purified by column chromatography over silica gel (gradient elution: n-heptane/EtOAc from 1:0 to 0:1). The fractions containing the product were collected and the solvent was evaporated to afford the title compound 43. Off-white powder. Yield: 80% (2 steps, 856 mg, 2.46 mmol). ¹H NMR (400 MHz, CDCl₃): δ 8.64-8.65 (m, 1H), 7.18 (d, J= 3.7 Hz, 1H), 6.61 (d, J= 3.7 Hz, 1H), 5.10 (dd, J= 6.3, 3.1 Hz, 1H), 4.96 (td, J= 6.8, 3.1 Hz, 1H), 4.82 (quin, J= 2.3 Hz, 1H), 4.78 (quin, J= 2.3 Hz, 1H), 4.69 (d, J= 6.5 Hz, 1H), 3.18 (dd, J= 15.5, 2.4 Hz, 1H), 2.75 (dd, J= 15.3, 2.6 Hz, 1H), 2.48-2.58 (m, 2H), 2.33-2.44 (m, 2H), 1.55 (s, 3H), 1.35 ppm (s, 3H). ¹³C NMR (101 MHz, CDCl₃): δ 152.3, 150.6, 143.4, 127.5, 117.9, 112.7, 107.0, 99.8, 85.7, 84.9, 61.5, 43.0, 42.3, 42.2, 38.3, 26.6, 24.7 ppm. LC-MS (method 3, ESI+) *m/z*: [M+H]⁺ Found 346.2 (Rt: 2.32 min).

Synthesis of 7-((3a'R,6'R,6a'S)-2',2'-Dimethyl-3methylenetetrahydrospiro[cyclobutane-1,4'cyclopenta[d][1,3]dioxol]-6'-yl)-7H-pyrrolo[2,3-d]pyrimidin-4amine 39. 4-Chloro-7-((3a'R,6'R,6a'S)-2',2'-dimethyl-3methylenetetrahydrospiro-[cyclobutane-1,4'-

cyclopenta[d][1,3]dioxol]-6'-yl)-7H-pyrrolo[2,3-d]pyrimidine 43 (850 mg, 2.46 mmol, 1.00 eq) was dissolved in 1,4-dioxane (25 ml) and NH_3 (aq. 28%, 60 ml) was added. The mixture was heated in a sealed pressure reactor to 100°C for 24 hours. Then, the solution was concentrated to a minimal volume in vacuo and coevaporated twice with toluene. The residue was purified via column chromatography over silica gel (gradient elution: CH₂Cl₂: MeOH from 1:0 to 4:1). Fractions containing the product were combined to afford the title compound 39. White solid. Yield: 98% (790 mg, 2.41 mmol). ¹H NMR (400 MHz, CDCl₃): δ 8.33 (s, 1H), 6.89 (d, J= 3.7 Hz, 1H), 6.34 (d, J= 3.5 Hz, 1H), 5.17 (br s, 2H), 5.08 (dd, J= 6.4, 2.9 Hz, 1H), 4.92 (td, J= 6.7, 2.9 Hz, 1H), 4.80 (quin, J= 2.4 Hz, 1H), 4.76 (quin, J= 2.4 Hz, 1H), 4.67 (d, J= 6.4 Hz, 1H), 3.16 (dq, J= 15.5, 2.4 Hz, 1H), 2.67-2.78 (m, 1H), 2.45-2.55 (m, 2H), 2.29-2.44 (m, 2H), 1.35 ppm (s, 3H). ¹³C NMR (101 MHz, CDCl₃): δ 156.6, 151.7, 150.5, 143.8, 123.0, 112.4, 106.8, 103.5, 97.6, 85.9, 85.1, 60.8, 43.1, 42.5, 42.3, 38.3, 26.6, 24.7 ppm. LC-MS (method 2, ESI+) m/z: [M+H]+ Found 327.3 (Rt: 0.89 min).

 Synthesis
 of
 (2S,4r,5R,6S,7R)-7-(4-amino-7Hpyrrolo[2,3-d]pyrimidin-7-yl)-2-((3-chloroquinolin-7yl)methyl)spiro[3.4]octane-5,6-diol 40a and (2R,4s,5R,6S,7R)-7-(4-amino-7H-pyrrolo[2,3-d]pyrimidin-7-yl)-2-((3

chloroquinolin-7-yl)methyl)spiro[3.4]octane-5,6-diol 40b. 7-

((3a'R,6'R,6a'S)-2',2'-Dimethyl-3-

methylenetetrahydrospiro[cyclobutane-1,4'cyclopenta[d][1,3]dioxol]-6'-yl)-7H-pyrrolo[2,3-d]pyrimidin-4amine 39 (52.0 mg, 0.16 mmol, 1.00 eq) was dissolved in 9borabicyclo[3.3.1]nonane (0.5M in THF, 1.91 ml, 0.96 mmol, 6.00 eq) at room temperature. The mixture was stirred for 30 minutes. Subsequently, potassium phosphate (271 mg, 1.28 mmol, 8.00 eq) dissolved in water (0.50 ml, 27.7 mmol, 173 eq) was degassed with nitrogen for 10 minutes and added to the reaction mixture. The solution was stirred for 10 minutes at room temperature while degassing and 7-bromo-3-chloroquinoline (49.8 mg, 0.207 mmol, 1.30 eq) and 1,1'-bis(di-tert-butylphosphino)ferrocene palladium dichloride (15.7 mg, 0.03 mmol, 0.15 eq) dissolved in THF (2.00 ml) was added to the mixture. Degassing with nitrogen was continued for 15 minutes before the mixture was heated to 70°C. After 2 hours, the dark brown solution was cooled to room temperature, diluted with EtOAc (90 ml), washed with NH4OH (25% in H₂O, 2x 30 ml) and brine (2x 30 ml). The organic layer was dried (MgSO₄), filtered and the filtrate was concentrated in vacuo. The residue (78.1 mg) was dissolved in EtOH (2 ml) and HCI (aq. 1M, 3 ml) was added. The mixture was stirred at room temperature for 1 hour. The solution was diluted with water (20 ml), frozen and lyophilized to afford a solid residue. A purification was performed via Prep SFC (Stationary phase: Chiralcel Diacel OJ 20 x 250 mm, Mobile phase: CO₂, EtOH + 0.4 *i*PrNH₂). Fractions containing the products were combined to afford the title compounds 40a and 40b. 40a: White solid. Yield: 21% (2 steps, 10.5 mg, 0.02 mmol). ¹H NMR (400 MHz, DMSO-d₆): δ 8.84 (d, J= 2.6 Hz, 1H), 8.51 (d, J= 2.4 Hz, 1H), 8.00 (s, 1H), 7.89 (d, J= 8.4 Hz, 1H), 7.82 (s, 1H), 7.52 (dd, J= 8.4, 1.5 Hz, 1H), 7.15 (d, J= 3.5 Hz, 1H), 6.86 (s, 2H), 6.52 (d, J= 3.5 Hz, 1H), 4.79-4.85 (m, 1H), 4.75 (dd, J= 10.1, 5.5 Hz, 2H), 4.17-4.24 (m, 1H), 3.71 (t, J= 4.8 Hz, 1H), 2.90 (d, J= 7.7 Hz, 2H), 2.53-2.67 (m, 1H), 2.27-2.40 (m, 1H), 2.14 (dd, J= 11.2, 8.8 Hz, 1H), 2.00-2.10 (m, 1H), 1.91 (dd, J= 13.2, 8.6 Hz, 1H), 1.85 (dd, J= 10.8, 8.6 Hz, 1H), 1.73 ppm (ddd, J= 11.2, 7.9, 3.5 Hz, 1H). ¹³C NMR (101 MHz, DMSO-d₆): δ 157.9, 151.7, 150.3, 149.5, 146.5, 143.8, 134.4, 129.9, 127.7, 127.7, 127.2, 127.0, 122.8, 103.2, 99.1, 78.4, 76.0, 60.0, 42.8, 42.7, 41.7, 38.5, 34.7, 30.8 ppm. HRMS (ESI⁺) *m/z*: [M+H]⁺ Calcd. for C24H25CIN5O2 450.1697; Found 450.1693 (Rt: 8.33 min). 40b: White powder. Yield: 26% (2 steps, 13.2 mg, 0.03 mmol). ¹H NMR (400 MHz, DMSO-d₆): δ 8.84 (d, J= 2.4 Hz, 1H), 8.51 (d, J= 2.2 Hz, 1H), 8.01 (s, 1H), 7.89 (d, J= 8.6 Hz, 1H), 7.82 (s, 1H), 7.52 (dd, J= 8.4, 1.3 Hz, 1H), 7.11 (d, J= 3.5 Hz, 1H), 6.87 (s, 2H), 6.54 (d, J= 3.5 Hz, 1H), 4.73-4.88 (m, 3H), 4.23-4.32 (m, 1H), 3.78 (t, J= 4.1 Hz, 1H), 2.89 (d, J= 7.0 Hz, 2H), 2.51-2.58 (m, 1H), 2.42-2.48 (m, J= 3.7 Hz, 1H), 2.29 (dd, J= 13.6, 10.3 Hz, 1H), 2.09-2.18 (m, 1H), 1.76-1.91 (m, 2H), 1.54 ppm (br dd, J= 10.5, 8.0 Hz, 1H). ¹³C NMR (101 MHz, DMSO-d₆): δ 157.9, 151.7, 150.5, 149.5, 146.5, 143.8, 134.4, 129.9, 127.7, 127.2, 127.0, 122.4, 103.2, 99.1, 77.9, 76.2, 59.3, 43.1, 43.0, 42.4, 40.8, 34.5, 30.7 ppm. HRMS (ESI⁺) *m*/*z*: [M+H]⁺ Calcd. for C₂₄H₂₅ClN₅O₂ 450.1697; Found 450.1692 (Rt: 8.31 min). **Computational Details**

The reduction of 4'-spirocyclobutanones 17 and 19 with the LiAlH₄ reducing agent was investigated using the Gaussian quantum chemistry package (version G09.D01).^[27] In line with our previous works on the facial selectivity of hydride reductions on 2cyclohexanones^[28] substituted and 3-substituted cyclobutanones,^[15] all quantum-chemical calculations were

performed using a two-step computational protocol. In our recent benchmark of DFT approximations,^[28] single-point energy calculations with the double hybrid B2PLYP-D3^[29] functional on the ωB97X-D^[30] optimized geometries were shown to provide very accurate transition state energies with respect to the canonical golden-standard CCSD(T) energies, whilst providing cis:trans ratios in good agreement with the experimental observations.

All the transition states were optimized and characterized by means of harmonic-vibrational-frequency calculations using a large "ultrafine" integration grid at the ω B97X-D/cc-pVDZ^[31] level of theory. The characteristic single negative eigenvalue of the Hessian matrix, coinciding with a transition state, was identified as the transfer of the hydride towards the carbonyl functionality. The thermodynamic corrections to the enthalpy (ΔH) and Gibbs free energy (ΔG) were computed at 1 atm for both room (298.15 K) and reaction (195.15 K) temperature. Subsequently, singlepoint energy calculations using the double hybrid B2PLYP-D3 functional and aug-cc-pVTZ^[32] basis set were performed on the ωB97X-D geometries. The DFT-D3 empirical dispersion correction of Grimme together with the Becke-Johnson damping function was included since it improves the description of noncovalent interactions and reaction barrier heights.^[33] Implicit solvent effects for THF were assessed on the gas-phase geometries by means of the polarizable continuum model with radii and non-electrostatic terms from Truhlar and co-workers' SMD model^[34] at the same level of theory. The stereoselectivity of the reductions (cis:trans ratios) was determined based on the Gibbs free energies of the different transition states (ΔG^{\neq}) using a Maxwell-Boltzmann distribution at 298.15 K and 195.15 K.

The effect of noncovalent interactions on the stereoselectivity was established using the noncovalent interaction (NCI) index,[17] as implemented in the NCIPLOT program,[35] starting from the wB97X-D/cc-pVDZ wavefunctions of the optimized geometries of the transition states. The NCI index allows to visualize the noncovalent interactions by plotting the reduced density gradient (s) versus the electron density (ρ) multiplied by the sign of the second eigenvalue of the electron density Hessian matrix [sign(λ_2)]. The latter is used to distinguish between attractive [sign(λ_2) < 0] and repulsive [sign(λ_2) > 0] interactions.^[17] A very important tool consists of visualizing the reduced gradient isosurfaces in real space. The value of the sign(λ_2) ρ is used to color the different isosurfaces using a RGB (red-blue-green) scale: red isosurfaces indicate repulsive interactions, blue stands for attractive interactions and green for very weak van der Waals-type interactions.

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Keywords: Carbocyclic nucleosides • cycloaddition • enzymatic reductions

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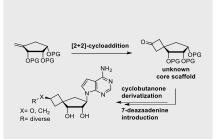
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Entry for the Table of Contents

FULL PAPER

A novel class of spirocarbocyclic scaffolds and derived nucleoside analogues is described. Using an exo-methylene building block, the [2+2] cycloaddition of dichloroketene affords a modular spirocyclobutanone scaffold structure in a key step. The reactivity evaluation of this core building block resulted in the introduction of versatile functional group handles for derivatization of the cyclobutanone ring. As a proof of concept, we showed that the corresponding nucleoside analogues could be prepared in an efficient manner, which have potential applications in medicinal chemistry.



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Synthesis and reactivity of novel spirocarbocycles as scaffolds for new nucleoside analogues